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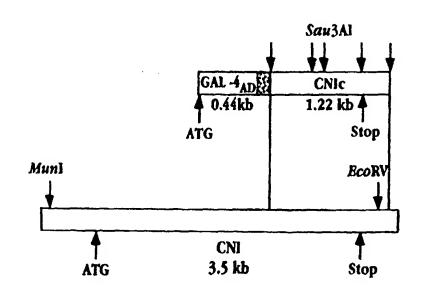
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### (57) Abstract

An identification and characterization of a calcineurin interacting (CNI) protein effective to enhance immunosuppressive effects calcineurin-targeted of immunosuppressants by potentiating an interaction of an immunophilin with calcineurin is described herein. One embodiment of the invention is the CNI polypeptide encoded by the CNI gene of Saccharomyces cerevisiae. Polynucleotides encoding a CNI protein are also described. Also described are yeast cells carrying mutations in the CNI gene. Further, a method of identifying a small molecule immunosuppressant compound is described. The methods include the use of a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains a subunit of calcineurin, and the other



of two fusion hybrid proteins contains an immunophilin.

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# CALCINEURIN INTERACTING PROTEIN COMPOSITIONS AND METHODS

PCT/US95/13580

### Field of the Invention

The present invention relates to compounds affecting the function of calcineurin, particularly interactions of calcineurin with immunosuppressant drugs.

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# 30 Background of the Invention

The immune system functions as the body's major defense against diseases caused by invading organisms. This complex system fights disease by killing invaders such as bacteria, viruses, parasites or cancerous cells while leaving the body's normal tissues unharmed. The immune system's ability to distinguish the body's normal tissues, or self, from foreign or

cancerous tissue, or non-self, is an essential feature of normal immune system function. A second essential feature is memory, the ability to remember a particular foreign invader and to mount an enhanced defensive response when the previously encountered invader returns. The loss of recognition of a particular tissue as self and the subsequent immune response directed against that tissue produce serious illness.

An autoimmune disease results from the immune system attacking the body's own organs or tissues, producing a clinical condition associated with the destruction of that tissue. An autoimmune attack directed against the joint lining tissue results in rheumatoid arthritis; an attack against the conducting fibers of the nervous system results in multiple sclerosis. The autoimmune diseases most likely share a common pathogenesis and the need for safe and effective therapy. One type of therapy that has been employed in combating autoimmune disease is treatment with immunosuppressant drugs, such as cyclosporin A, FK506 and rapamycin. While the treatments are often effective, the drugs typically have undesirable side effects, including neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Many of these side effects are due to the drugs' action on cells other than those of the immune system.

In addition to their use in treating autoimmune conditions, immunosuppressive agents have also been used in treating or preventing transplantation rejection. Organ transplantation involving human organ donors and human recipients (allogeneic grafts), and non-human primate donors and human recipients (xenogeneic grafts), has received considerable medical and scientific attention (e.g., Roberts, 1989; Platt, 1990). To a great extent, this effort has been aimed at eliminating, or at least reducing, the problem of rejection of the transplanted organ. In the absence of adequate immunosuppressive therapy, the transplanted organ is destroyed by the host immune system.

Presently, the most commonly used agents for preventing transplant rejection include corticosteroids, cytotoxic drugs that specifically inhibit T cell activation such as azathioprine, immunosuppressive drugs such as cyclosporin A, and specific antibodies directed against T lymphocytes or surface receptors that mediate their activation (Briggs, 1991; Kennedy, 1983; Storb, 1985; Storb, 1986). All of these drug therapies are limited in effectiveness, in part because the doses needed for effective treatment of transplant rejection may increase the patient's susceptibility to infection by a variety of opportunistic invaders, and in part because of direct toxicity and other side effects.

Cyclosporin A, currently the most effective and most commonly used agent, is significantly toxic to the kidney. This nephrotoxicity limits the quantity of drug that can be

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safely given. The physician is frequently forced to administer sub-optimal doses of the drug because of this toxicity. A preparation capable of potentiating the action of immunosuppressive agents such as cyclosporin A on the immune system, thus allowing the administration of lower doses of drug, would be of considerable value in reducing the morbidity and mortality associated with transplantation.

### Summary of the Invention

In one embodiment, the present invention includes polypeptide compositions effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin. The present invention includes the isolation and characterization of a calcineurin interacting protein, CNI, having these properties. Also disclosed herein are methods for the isolation and characterization of further CNI-related sequences and sequences of CNI-variants. The amino acid sequences presented as SEQ ID NO:2 and SEQ ID NO:5 are exemplary of the polypeptides of the present invention.

The present invention also includes a CNI polypeptide fragment that interacts specifically with the "A" subunits of calcineurin (CNA1 and CNA2), but not with calcineurin "B" subunit (CNB1). In one embodiment, this fragment has an amino acid sequence of between 15 and 915 amino acids in length, for example, the c-terminal 306 amino acids of the CNI protein (CNIc).

Included aspects of the invention are an CNI polypeptide; a recombinant CNI polypeptide; and a fusion polypeptide comprised of an CNI polypeptide. Exemplary fusion proteins include fusions to  $\beta$ -galactosidase.

The invention further includes isolated nucleic acid sequences encoding the above described polypeptides and polypeptide fragments. Exemplary nucleic acid sequences include the sequences presented as SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:6. The present invention includes CNI-encoding genomic polynucleotides, cDNAs thereto and complements thereof. With respect to polynucleotides, some aspects of the invention include: a purified CNI-encoding genomic polynucleotide; CNI polypeptide-encoding RNA and DNA polynucleotides; recombinant CNI polypeptide-encoding polynucleotides; a recombinant vector including any of the above recombinant polynucleotides, and a host cell transformed with any of these vectors. Another aspect of the invention is a polynucleotide probe for CNI polypeptide-encoding sequences.

Portions of a CNI-polypeptide coding sequences are effective as probes to isolate variants coding sequences which occur naturally, or to determine the presence of such coding

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sequences in nucleic acid samples. Such probes include hybridization screening probes and polymerase chain reaction amplification primers specific for CNI-polypeptide coding sequences. Homologues of CNI may be isolated from a number of sources, such as other types of yeast cells (e.g., Schizosaccharomyces) or mammalian cells (e.g., human).

Other aspects of the invention include: a recombinant expression system which incorporates an open reading frame (ORF) derived from CNI polypeptide-encoding sequences, wherein the ORF is linked operably to a control sequence which is compatible with a desired host, a cell transformed with the recombinant expression system, and a polypeptide produced by the transformed cell. Typically the expression system includes a vector having (a) a nucleic acid containing an open reading frame that encodes a CNI-polypeptide; and (b) regulatory sequences effective to express said open reading frame in a host cell. The regulatory sequence may include sequences useful for targeting or secretion of the CNI-polypeptide: such as a secretory signal recognized in yeast or bacterial expression systems.

The invention includes a method of recombinantly producing CNI-polypeptides. In the method, a recombinant expression system containing an open reading frame (ORF) having a polynucleotide sequence which encodes a CNI-polypeptide, where the vector is designed to express the ORF in the host, is introduced into suitable host cells. The host is then cultured under conditions resulting in the expression of the ORF sequence. The CNI-polypeptide sequences discussed above are examples of suitable CNI-polypeptides. Numerous vectors and their corresponding hosts are useful in the practice of this method of the invention, including, but not limited, to the vectors described herein for expression in yeast cells, and lambda gt11 phage vector and E. coli cells. Other host cells include insect and mammalian cell expression systems.

The invention also includes purified antibodies that are immunoreactive with a CNI-polypeptide. The antibodies may be polyclonal or monoclonal. Antibodies that are specifically immunoreactive with CNI-polypeptides may be useful for the isolation of CNI-polypeptide homologues from other cell type sources (e.g., mammalian).

The present invention also includes, a method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed where one of two fusion hybrid proteins in the cell contains a subunit of calcineurin, and the other of two fusion hybrid proteins contains an immunophilin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. In one embodiment, the method is carried out using yeast cells, where one of the two

fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells (including human cells). The subunit may, for example, be yeast calcineurin subunit CNA1 or CNA2, or human calcineurin subunit "A". The immunophilin can, for example, be cyclophilins or FK506-binding proteins (e.g., FKBP12) typically from a homologous cell source.

Also included in the present invention is another method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed, wherein one of two fusion hybrid proteins in a cell contains an "A" subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI polypeptide. The cell preferably, but not necessarily, also contains a vector construct causing overexpression, or increased expression, of a "B" subunit of calcineurin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. This method is used to identify compounds (like FK506) that potentiate the interaction between CNI and CNA1. In one embodiment, the method is carried out using yeast cells, where one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells 20 (including human cells). The subunits may, for example, be calcineurin subunit A1 or A2. The CNI polypeptide may also be from any source (e.g., yeast or human), and may be only a fragment of a complete CNI polypeptide (such as a c-terminal fragment). An exemplary cterminal fragment of CNI is CNIc.

Further, included in the present invention, is a yeast cell carrying a mutation in the naturally-occurring copy of CNI, where the mutation prevents expression of a functional CNI protein from the genomic copy. Embodiments of this aspect of the present invention include deletion mutations within the coding region of the CNI gene, deletion of regulation regions of the CNI gene, and non-sense or mis-sense mutations in the CNI gene. Yeast cells having such mutations are useful, for example, in a method of identifying proteins of similar function to CNI. In one embodiment, a hybrid interaction screen is set up in a cell with a CNI deletion and a GAL4 protein binding domain-CNA fusion and a GAL4 activation domain-immunophilin fusion. Expression libraries are then screened to identify clones encoding proteins that potentiate an interaction of an immunophilin with calcineurin. This screen will identify CNI-coding sequences as well as other proteins with a similar function.

In a related embodiment, a yeast cell with a CNI deletion is used to identify CNI homologues (e.g., from other organisms, such as human) using a complementation assay or screen. Expressionlibraries (e.g., human lymphocyte expression libraries) are transformed into cells with a CNI deletion, and transformants are selected on their ability to complement the function of yeast CNI. An exemplary assay for selecting such transformants is exposure to hygromycin B. Cells which become more sensitive to hygromycin B following transformation are further analyzed to determine if the plasmid with which they were transformed contains an insert homologous to yeast CNI, or encoding a polypeptide with similar function to CNI.

The invention also includes a yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where the mutation prevents expression of a functional calcineurin-interacting polypeptide from the genomic copy. The mutation may be a null mutation, such as described in Example 8 below, or a different type of mutation, e.g., a nonsense or missense mutation. Nonsense and missense mutations may be generated using standard methods.

These and other objects and features of the invention will be more fully appreciated when the following detailed description of the invention is read in conjunction with the accompanying drawings.

# Brief Description of the Figures

Figures 1A and 1B present schematic diagrams of sequences encoding the c-terminal portion of CNI (CNIc) fused to GAL-4 activation domain (GAL-4AD) (Fig. 1A), and sequences encoding CNI (Fig. 1B).

Figure 2A presents data from a  $\beta$ -galactosidase ( $\beta$ -gal) assay to detect the interaction of CNIc with the A1 subunit of calcineurin (CNA1), A2 subunit of calcineurin (CNA2), GAL-4 binding domain (G4BD) and lamin C. A labeled schematic diagram corresponding to the data shown in Fig. 2A is presented in Fig. 2B to facilitate reference to individual groups of colonies.

Figure 3A presents data from a  $\beta$ -gal assay to detect the interaction of CNIc with CNA1 $\Delta$ C, CNA2 $\Delta$ C and CNB1. A labeled schematic diagram corresponding to the data shown in Fig. 3A is presented in Fig. 3B.

Figures 4A and 4C present data from  $\beta$ -gal assays to evaluate the effects of FK506 and the deletion of CNB1 on the interactions of CNIc with CNA and CNA $\Delta$ C. Labeled schematic diagrams corresponding to the data shown in Figs. 4A and 4C are presented in Figs. 4B and 4D, respectively.

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Figures 5A and 5C present data from  $\beta$ -gal assays to evaluate the effects of FK506 and the overexpression of CNB1 on the interactions of CNIc with CNA and CNA $\Delta$ C. Labeled schematic diagrams corresponding to the data shown in Figs. 5A and 5C are presented in Figs. 5B and 5D, respectively.

Figures 6A, 6C and 6E present data from  $\beta$ -gal assays to evaluate the effects of FK506, cyclosporin A (CsA), and the deletion of CNB1 on the interaction of CNIc with CNA and CNA $\Delta$ C. Labeled schematic diagrams corresponding to the data shown in Figs. 6A, 6C and 6E are presented in Figs. 6B, 6D and 6F, respectively.

Figures 7A, 7C, 7E and 7G present data from β-gal assays to evaluate the effects of FK506, cyclosporin A (CsA), rapamycin and the overexpression of CNB1 on the interaction of CNIc with CNA and CNAΔC. Labeled schematic diagrams corresponding to the data shown in Figs. 7A, 7C, 7E and 7G are presented in Figs. 7B, 7D, 7F and 7H, respectively.

Figure 8A presents data from a  $\beta$ -gal assay to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNA. A labeled schematic diagram corresponding to the data shown in Fig. 8A is presented in Fig. 8B.

Figures 9A and 9C present data from  $\beta$ -gal assays to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNB1. Labeled schematic diagrams corresponding to the data shown in Figs. 9A and 9C are presented in Figs. 9B and 9D, respectively.

Figure 10 presents an image of a protein blot of CNIc and CNA co-immunoprecipitate probed with anti-CNA2 antibody.

Figure 11 presents an image of a yeast RNA blot hybridized with a CNIc probe.

Figure 12 presents the DNA sequence of a 3.5 kb fragment of yeast chromosome 11 containing the coding sequence for a yeast CNI protein.

25 Figure 13 presents the amino acid sequence of a yeast CNI protein.

### Brief Description of the Sequences

SEQ ID NO:1 presents the nucleotide sequence of a Sau3AI fragment containing the coding sequence for CNIc.

SEQ ID NO:2 presents the amino acid sequence of CNIc encoded by SEQ ID NO:1.

SEQ ID NO:3 presents the coding sequence presented in SEQ ID NO:1.

SEQ ID NO:4 presents the nucleotide sequence of a gene encoding a complete CNI protein.

SEQ ID NO:5 presents the amino acid sequence encoded by SEQ ID NO:4.

SEQ ID NO:6 presents the coding sequence presented in SEQ ID NO:4.

SEQ ID NO:7 presents the nucleotide sequence of PCR primer CNI-PCR-A.

SEQ ID NO:8 presents the nucleotide sequence of PCR primer CNI-PCR-B.

SEQ 1D NO:9 presents the nucleotide sequence of a gene encoding the yeast CNA1 subunit of calcineurin.

SEQ ID NO:10 presents the amino acid sequence encoded by SEQ ID NO:9.

SEQ ID NO:11 presents the nucleotide sequence of a gene encoding the yeast CNA2 subunit of calcineurin.

SEQ ID NO:12 presents the amino acid sequence encoded by SEQ ID NO:11.

SEQ ID NO:13 presents the nucleotide sequence of a gene encoding the yeast CNB1 subunit of calcineurin.

SEQ ID NO:14 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:15 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:16 presents the coding sequence presented in SEQ ID NO:13.

15 SEQ ID NO:17 presents the amino acid sequence encoded by SEQ ID NO:16.

SEQ ID NO:18 presents a nucleotide sequence encoding CNA1 \( \Delta \)c.

SEQ ID NO:19 presents the amino acid sequence encoded by SEQ ID NO:18.

SEQ ID NO:20 presents a nucleotide sequence encoding CNA2Δc.

SEQ ID NO:21 presents the amino acid sequence encoded by SEQ ID NO:20.

SEQ 1D NO:22 presents the nucleotide sequence of PCR primer G4-PCR-A.

SEQ 1D NO:23 presents the nucleotide sequence of PCR primer G4-PCR-B.

# Detailed Description of the Invention

### I. DEFINITIONS

A "calcineurin-targeted immunosuppressant" is a compound that possesses in vivo immunosuppressive activity, and that interacts with an immunophilin to form a complex which is capable of inhibiting calcineurin.

"Interacting proteins" are proteins capable of specifically binding to one another, or associating with one another, in a cell or *in vitro*.

A calcineurin interacting (CNI) protein or polypeptide is a protein or polypeptide that is effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin. Preferably, a CNI protein or polypeptide is a protein or polypeptide having an amino acid sequence that is homologous to the sequence presented herein as SEQ ID NO:5.

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"Substantially isolated" is used in several contexts and typically refers to the at least partial purification of a CNI protein or polypeptide fragment away from unrelated or contaminating components ( $\epsilon$ .g., cytoplasmic contaminants and heterologous proteins). Methods and procedures for the isolation or purification of compounds or components of interest are described below ( $\epsilon$ .g., affinity purification of fusion proteins and recombinant production of CNI polypeptides).

In the context of the present invention, the phrase "nucleic acid sequences," when referring to sequences which encode a protein, polypeptide, or peptide, is meant to include degenerative nucleic acid sequences which encode homologous protein, polypeptide or peptide sequences as well as the disclosed sequence.

Two nucleic acid fragments are considered to have "homologous" sequences if they are capable of hybridizing to one another (i) under typical hybridization and wash conditions, as described, for example, in Sambrook, et al., pages 320-328, and 382-389, or (ii) using reduced stringency wash conditions that allow at most about 25-30% basepair mismatches, for example: 2 × SSC, 0.1% SDS, room temperature twice, 30 minutes each; then 2 × SSC, 0.1% SDS, 37°C once, 30 minutes; then 2 × SSC, room temperature twice, 10 minutes each. Preferably, homologous nucleic acid strands contain 15-25% basepair mismatches, even more preferably 5-15% basepair mismatches. These degrees of homology can be selected by using wash conditions of appropriate stringency for identification of clones from gene libraries (or other sources of genetic material), as is well known in the art.

Two amino acid sequences or two nucleotide sequences (in an alternative definition for homology between two nucleotide sequences) are considered homologous (as this term is preferably used in this specification) if they have an alignment score of >5 (in standard deviation units) using the program ALIGN, typically default mutation gap matrix and gap penalty (Dayhoff). The two sequences (or parts thereof) are more preferably homologous if their amino acids are greater than or equal to 40% using the ALIGN program mentioned above.

# II. OVERVIEW OF INVENTION

Experiments performed in support of the present invention demonstrate the identification and isolation of the nucleic acid sequence encoding a calcineurin interacting (CNI) protein. Further experiments performed in support of the present invention characterize the CNI protein, as well as a polypeptide containing only the c-terminal 306 amino acids of the CNI protein (CNIc). The experiments demonstrate that CNIc interacts specifically with the "A" subunits of calcineurin (CNA1 and CNA2), but not with calcineurin "B" subunit (CNB1).

The experiments also demonstrate that CNIc does not interact directly with FK506 binding protein (FKBP; Schreiber,  $et\ al.$ ; with or without FK506), GAL4 binding domain (G4<sub>bD</sub>) or lamin C. The experiments also demonstrate that CNIc interacts with C-terminally truncated forms of CNA (CNA $\Delta$ C), which have lost their autoinhibitory domains, though the interaction is somewhat weaker that with full length CNA proteins.

Additional experiments show that the interaction between CNIc and CNA is enhanced when CNB1 is deleted, and diminished when CNB1 is overexpressed, that the interaction between CNIc and CNA or CNA $\Delta$ C is markedly enhanced by FK506 and by Cyclosporin A (CsA), but not rapamycin, and that overexpression of a full-length CNI protein enhances the interaction between CNA and FKBP (detectable only in the presence of FK506).

Additional experiments conducted in support of the present invention demonstrate that overexpression of the full-length CNI has no detectable effect on the interaction between CNB1 and CNA, and that in the presence of FK506 or CsA, overexpression of CNB1 no longer inhibits the interaction of CNIc with CNA.

It was also found that CNI deletion mutants are viable, both in wild-type and CN-deletion backgrounds, and that CNI deletion mutants in a CN-deletion background are more resistant to hygromycin B than normal CN-deletion mutants.

Co-immunoprecipitation experiments demonstrate that CNIc and CNA co-immunoprecipitate in the presence of FK506, and protein blot experiments show that CNI is expressed at low levels in vivo. RNA blot experiments show that CNI is encoded by a single message approximately 2.9 kb in length.

A comparison of the yeast CNI sequence with sequences present in nucleic acid and amino acid databases reveals no obvious homologous sequences have been identified in other organisms.

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# III. CALCINEURIN

Experiments performed in support of the present invention were designed to identify polypeptides capable of interacting with calcineurin. Calcineurin (also called phosphoprotein phosphatase 2B or PP2B), has been characterized from many different tissues and organisms (Klee, et al.). It is a heterodimer of two subunits, of which the "A" subunit is about 61 kD in weight, possesses catalytic activity and also contains the association site for calmodulin. The "B" subunit contains four Ca2+ binding sites and activates the A subunit. Calcineurin has little enzymatic activity, even in the presence of Ca2+ and only becomes fully active when associated with calmodulin (Cvert).

Two A subunits (CNA1 and CNA2; Cyert, et al., 1991) and one B subunit (CNB1, Cyert, et al., 1992) have been cloned in yeast. Either CNA1 or CNA2 may associate with CNB1 to form a functional calcineurin heterodimer. Multiple isotypes of the A subunit have been cloned from a variety of organisms and are highly conserved (Klee, et al.). In particular, calcineurin subunits have been cloned from human tissue (see reviews by, for example, Klee, et al., and Guerini, et al.).

### IV. IMMUNOSUPPRESSANT DRUGS

FK506, cyclosporin A (CsA) and rapamycin, derived from fungi, inhibit the activation of T-cells by antigens. The compounds have proven highly effective at suppressing mammalian immune systems in vivo. In particular, CsA therapy in clinical settings has dramatically increased the success rate of transplantation therapy.

It is now known that FK506 and CsA exert their immunosuppressive effects, in part, by inhibiting the transcriptional activation of the interleukin-2 (IL-2) gene, whereas rapamycin appears to function by inhibiting the response of T-cells to IL-2, presumably by inhibiting a transduction pathway mediated by the IL-2 receptor.

The molecular mechanism of FK506 and CsA immunosuppressive action involves a group of small, abundant intracellular proteins termed immunophilins, which bind with a high affinity to the immunosuppressants (Schreiber). At least two classes of immunophilins are known to exist. One class, termed cyclophilins, binds to CsA, while another class, the FK506-binding proteins (FKBPs) binds FK506 and rapamycin. Many immunophilin genes, from a variety of organisms, have been cloned, and appear to be highly conserved from simple eukaryotes to mammals.

It is believed that FK506 and CsA-induced immunosuppression is due to the binding of complexes, formed by binding of immunosuppressants FK506 and CsA bound to one of their respective immunophilins, to the catalytic subunit of calcineurin (Schreiber, et al., Liu, et al., Foor, et al., Weiss, et al.). The binding of such a complex to an (A) subunit inhibits activation of calcineurin by increased intracellular calcium, which in turn prevents calcineurin from activating transcription factor NF-AT. Since IL-2 is one of the genes controlled by NF-AT in T-cells, inhibition of the transcription factor inhibits the production of IL-2, resulting in immunosuppression (Clipstone, et al.).

FK506 and CsA are widely used in organ transplantation to prevent host rejection. However, both drugs are known to have many undesired side-effects such as neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Accordingly formulations effective to

increase a target cell's sensitivity to these drugs may be useful in alleviating some of the aforementioned side-effects. Specifically, CNI and its homologues or derivatives, administered at appropriate levels, may be able to increase the sensitivity of CN to FK506/CsA and reduce the necessary dosage thus reducing or eliminating the side-effects of these drugs.

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# V. Two Hybrid Protein Interaction Assays

Two hybrid protein interaction assay methods (two hybrid protein-protein interaction screens) provide a simple and sensitive means to detect the interaction between two proteins in living cells. The assays are based on the finding that most eukaryotic transcription activators are modular (e.g., Brent, et al.), i.e., that the activators typically contain activation domains that activate transcription, and DNA binding domains that localize the activator to the appropriate region of a DNA molecule.

The development of two hybrid protein interaction assays was made possible by the observation that the DNA binding domain does not need to be physically located on the same polypeptide as the activation domain (Ma, et al., Triezenberg, et al.), raising the possibility that transcription of reporter genes could be used as an assay to detect protein interactions.

The utility of two hybrid systems for detecting interactions between two interacting proteins was fully realized by the observation that protein interactions could be detected if two potentially-interacting proteins were expressed as fusions, or chimeras (Fields, et al.). A first fusion protein contains one of a pair of interacting proteins fused to a DNA binding domain, and a second fusion protein contains the other of a pair of interacting proteins fused to a transcription activation domain. The two fusion proteins are independently expressed in the same cell, and interaction between the "interacting protein" portions of the fusions reconstitute the function of the transcription activation factor, which is detected by activation of transcription of a reporter gene.

At least two different cell-based two hybrid protein-protein interaction assay systems have been used to assess binding interactions and/or to identify interacting proteins. Both employ a pair of fusion hybrid proteins, where one of the pair contains a first of two "interacting" proteins fused to a transcription activation domain of a transcription activating factor, and the other of the pair contains a second of two "interacting" proteins fused to a DNA binding domain of a transcription activating factor.

The yeast GAL4 two hybrid system (Fields, et al., Chien, et al., Durfee, et al., Bartel, et al.), utilized for experiments performed in support of the present invention, was developed

to detect protein-protein interaction based on the reconstitution of function of GAL4, a transcriptional activator from yeast, by activation of a GAL1-lacZ reporter gene.

Like several other transcription activating factors, GAL4 contains two distinct domains, a DNA binding domain and a transcription activation domain. Each domain can be independently expressed as a portion of a fusion protein composed of the domain, and a second, "bait" interacting protein. The two fusion proteins are then independently expressed together in a cell. When the two GAL4 domains are brought together by a binding interaction between the two "interacting" proteins, transcription of a reporter gene under the transcriptional control of GAL4 is initiated. The reporter gene typically has a promoter containing GAL4 protein binding sites (GAL upstream activating sequences, UAS<sub>0</sub>). Exemplary reporter genes are the GAL1-lacZ, and GAL1-HIS3 reporter genes used in experiments described herein.

A second two hybrid system, described in detail in Ausubel, et al., utilizes a native E. coli LexA repressor protein, which binds tightly to appropriate operators. A plasmid is used to express one of a pair of interacting proteins (the "bait" protein) as a fusion to LexA. The plasmid expressing the LexA-fused bait protein is used to transform a reporter strain of yeast, such as EGY48, that contains pSH18-34.

In this strain, binding sites for LexA are located upstream of two reporter genes. In the first reporter system, the upstream activation sequences of the chromosomal LEU2 generequired in the biosynthetic pathway for leucine (Leu)-are replaced in EGY48 with lexA operators, permitting selection for viability when cells are plated on medium lacking Leu. In the second reporter system, EGY48 harbors a plasmid, pSH18-34, that contains a lexA operator-lacZ fusion gene, permitting discrimination based on color when the yeast is grown on medium containing Xgal (Ausubel, et al.).

To screen a library with the LexA system, the library uses the inducible yeast GAL1 promoter to express proteins as fusions to an acidic domain ("acid blob") that functions as a portable transcriptional activation motif ("act"), and to other useful moieties. Expression of library-encoded proteins is induced by plating transformants on medium containing galactose (Gal), so yeast cells containing library proteins that do not interact specifically with the bait protein fail to grow in the absence of Leu. Yeast cells containing library proteins that interact with the bait protein form colonies within 2 to 5 days, and the colonies turn blue when the cells are streaked on medium containing Xgal. The plasmids are isolated and characterized by a series of tests to confirm specificity of the interaction with the initial bait protein. Those found to be specific are ready for further analysis (e.g., sequencing).

LexA and GAL4 each have different properties that should be considered when selecting a system. LexA is derived from a heterologous organism, has no known effect on the growth of yeast, possesses no residual transcriptional activity, can be used in GAL4\* yeast, and can be used with a Gal-inducible promoter. Because GAL4 is an important yeast transcriptional activator, experiments must be performed in gal4 yeast strains to avoid background from endogenous GAL4 activating the reporter system.

Both two hybrid systems have been successfully used for isolating genes encoding proteins that bind a target protein and as simple protein binding assays (e.g., Yang, et al., Gyuris, et al.), and both can be applied to methods of the present invention.

Both gene isolation and protein binding assay applications of the GALA system are described in Examples below.

# VI. SPECIFIC EMBODIMENTS

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Example 1 demonstrates application of an exemplary two hybrid protein-protein interaction screen (Materials and Methods, section D) to the screening of three pGAD yeast fusion libraries, carrying fusions between the transcription activating domain of yeast protein GAL4 (G4AD) and yeast genomic DNA Sav3Al fragments, in all three reading frames. The libraries are screened to identify polypeptides, encoded by the Sau3Al fragments, capable of interacting with catalytic (A) subunits of calcineurin, expressed as fusions with the GAL4 protein binding domain (GBT-CNA fusions).

Three sets of yeast cells harboring pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid and a GALA-activated LacZ reporter gene are each transformed with one of the three reading-frame libraries. Construction of the plasmids used is described in Materials and Methods, sections B and C. Cells transformed with a plasmid encoding a protein fusion capable of interacting with the CNA subunit fusion are selected using a  $\beta$ -galactosidase ( $\beta$ -gal) assay on plates containing the chromogenic substrate X-gal (Materials and Methods, section E). Results of the  $\beta$ -gal assay are confirmed using a growth assay (Materials and Methods, section F). False positives are eliminated by colony purification (re-streaking for single colonies), PCR experiments using GAL4 primers, and testing against a number of test fusions by  $\beta$ -gal assays on transformed haploid or mated diploid reporter strains.

A yeast clone encoding a polypeptide capable of specifically interacting with CNA polypeptide fusions is identified and sequenced. The sequence of the Sau3AI fragment is presented as SEQ ID NO:1. The coding sequence forming the open reading frame is presented as SEQ ID NO:3. The polypeptide encoded by the open reading frame is presented as SEQ

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ID NO:2. The open reading frame encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GAL4, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product. The clone is termed CNIc, with the lowercase "c" representing "cterminal".

Figure 1A shows a schematic representation of the nucleic acid sequence encoding the GAL4AD-CNIc fusion protein. The stippled portion between GAL4AD and CNIc represents a linker discussed in the Materials and Methods section, as well as in Example 1. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons. and Sau3Al restriction sites.

Example 1 further describes the identification of a \( \lambda \) clone encoding a full length sequence version of CNIc, termed CNI. The polypeptide encoded by the sequence is termed CNI protein. The clone is identified by hybridization screening of a panel of  $\lambda$  clones spanning the yeast genome using a 1.22 kb <sup>32</sup>P-labeled probe generated from CNIc.

Phage lysates of the  $\lambda$  clone are amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments. A 3.16 kb Munl/EcoRV fragment from the λ clone insert contains the coding sequence of CNI. The sequence of the 3.16 kb Munl/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4, and a schematic diagram of the sequence is shown in Figure 1B. This sequence contains the entire 2.75 kb 20 coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

A search of known DNA and protein sequences turns up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin interacting protein.

The methods referred to above may also be applied to the screening of, for example, a human cDNA library using an appropriate two-hybrid protein interaction screen. The "bait" protein in the interaction screen (e.g., the protein analogous to CNA1 in Example 1) may be of yeast origin (e.g., CNA1), but is preferably of human origin (e.g., a human calcineurin "A" subunit; Klee, et al.). The bait protein is expressed in the cell (e.g., a yeast cell) used for the 30 two hybrid interaction screen as a fusion to a domain of a transcription activating factor (e.g., the DNA binding domain of GAL4). The library may be a human DNA library in a vector (e.g., pGAD) effective to express library sequences as fusions to a complimentary domain of the transcription activating factor (e.g., the activation domain of GAL4). Libraries of human sequences can be derived from a number of sources including genomic DNA, such as yeast

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artificial chromosome (YAC) constructs carrying genomic human DNA, or cDNA generated from a variety of cell types (e.g., activated T-cells).

Example 2 details a \(\beta\)-gal assay to determine the specificity of binding of CNIc to subunits of calcineurin. Exemplary results are shown in Figure 2A. The legend for Fig. 2A 5 is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs (indicated in Example 2).

A comparison of the intensities of the blue  $\beta$ -gal reaction product indicates that CNIc interacts strongly with CNA1 (21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with 10 GAD-CNlc (26) show a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact with each other are used as a positive control for the assay (20). The data presented in Figure 2A show that CNIc interacts specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments, illustrated in Fig. 3A, is conducted using constitutivelyactive CNA subunits, as well as calcineurin subunit CNB1. As described in the Materials and Methods section below, CNA1\DC and CNA2\DC are each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A. The legend for Fig. 3A is shown in Figure 3B. The data show that GBT-A1\(\Delta\) and GAD-CNIc (28) gives a definite positive signal, while GBT-A2\(\Delta\) and 20 GAD-CNlc (29) is weaker, though still detectable above its background (i.e. GBT-A2ΔC and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) is not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 $\Delta$ C and GAD-B1 (34) and GBT-A2\(\Delta\) and GAD-B1 (35) give strong signals. The data presented in Figure 3A show that CNIc interacts specifically with CNA1 $\Delta$ C and CNA2 $\Delta$ C, but not with CNB1.

Example 3 details the effects of immunosuppressant drugs on binding of CNIc to calcineurin in B1<sup>nd</sup>, B1 Deletion and B1 Overproducing Yeast Strains. The yeast strains are assayed for  $\beta$ -gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments are performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 30 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data are shown in Figures 4A, 4C, 5A and 5C. The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Figs. 4A and 5A illustrate experiment

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performed without FK506, while experiments shown in Figs. 4C and 5C were performed with FK506.

The interactions of various combinations of proteins expressed by constructs indicated in Example 3 was studied in three yeast strains, one of which is null for the CNB1 subunit of calcineurin (Y153b; at 36-40), while the others (Y190; at 41-46 and Y526 at 47-51) are wild-type for CNA1, CNA2 and CNB1.

The data, shown in Fig. 4A (no added drugs), illustrate that deleting the endogenous host CNB1 gene potentiates, or enhances, interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1 $\Delta$ C and CNA2 $\Delta$ C. Comparison of corresponding colonies in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA $\Delta$ C interactions. The drug enhances interactions under all except control conditions. The effect is most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhances, or potentiates CNIc-CNA/CNAAC interactions under conditions where the CNB1 subunit is overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNAAC interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) have reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhances, or potentiates interactions in all colonies, except the negative controls (56).

The data presented in Figures 4A, 4C, 5A and 5C demonstrate that the interaction of CNIc with CNA and CNA\(Delta\) is markedly enhanced by FK506. The interaction is also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction is overcome by the stimulatory effect of FK506.

Stated another way, inclusion of a small molecule immunosuppressant (FK506) potentiates an interaction between two fusion hybrid proteins, where one of the two proteins contains an (A) subunit of calcineurin, and the other protein contains a CNI polypeptide. The potentiation is particularly strong when the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell (e.g., the expresses B1/YEp352).

In the present case, a yeast cell is modified to cause overexpression of a "B" subunit of calcineurin (CNB1) by transforming the cell with B1/YEp352 (construction described below). A cell may be modified to cause overexpression of a "B" subunit of calcineurin in other ways as well, such as, for example, transformation with other types of expression vectors encoding a "B" subunit of calcineurin, or treatment with a substance that upregulates a promoter controlling expression of an endogenous (B) subunit of calcineurin.

In light of the effects of FK506 on CNIc-CNA/CNAAC interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A is also effective to enhance interaction of CNIc with CNA and CNA\(Delta\)C. Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through calcineurin (Cyert).

Similarly, data shown in Figures 7A and 7C support data in Figures 5A and 5C, and 10 further, demonstrate that there is no detectable interaction between FK506 binding protein (FKBP) and CNIc. Results shown in Fig. 7E demonstrate that cyclosporin A has a similar effect to FK506 in cells overexpressing CNB1 — that is, it enhances the interactions between CNIc and CNA/CNAAC.

Taken together, the data presented in Figs. 6A, 6C, 6E 7A, 7C, 7E and 7G show that like FK506, cyclosporin A (CsA), but not rapamycin, enhances the interaction of CNIc with CNA and CNAΔC, and that CNIc doesn't interact with FKBP with or without FK506.

Example 4 describes experiments to assess effects of CNIc on FKBP/FK506 binding to calcineurin. Figure 8A presents exemplary data from studies to assess the effect of CNI on FKBP-mediated FK506 interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 show no detectable interaction (84). In the presence of FK506, however, the proteins interact (85), presumably because FK506 forms a complex with FKBP, which then binds CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI has no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiates, or enhances the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

Experiments illustrated in Figures 9A and 9B demonstrate that FK506 has little or no effect on the binding of CNA to CNB1. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively. The data in Fig. 9A show that overexpression of the full-length CNI

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clone markedly enhances the FK506-dependent interaction of FKBP with CNA, although it doesn't affect the interaction between CNA and CNB1.

Example 5 presents co-immunoprecipitation of CNIc (carrying an HA epitope tag) and CNA. Immunoprecipitation is carried out with anti-HA monoclonal antibody and the immune complex, resolved by SDS-PAGE, is detected with anti-CNA2 polyclonal antibody and visualized with goat anti-rabbit antibody using the "ECL" method (Amersham, Arlington Heights, IL). The results, shown in Figure 10, demonstrate that CNIc is capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. Similar methods may be employed to isolate a CNI analog from other cell sources, including mammalian (specifically human).

Example 6 describes yeast RNA blots hybridized with a CNIc probe. Exemplary data are shown in Figure 11. A single message of approximately 2.9 kb is detected. The data indicate that CNI is an expressed gene encoding a 2.9 kb message in yeast.

Example 8 details the construction of cni null mutants. The null mutants are employed to assess if CNI is required for viability in yeast, and to test hygromycin B sensitivity. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains can survive, but that CNI deletions render a host more resistant to hygromycin B. The effect is particularly pronounced in both MCY300-1 (cna1 cna2) and DD12 (cnb1), suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B.

It will be understood that all of the above methods and experimental manipulations are amenable to being done with interacting polypeptides from organisms other than yeast. In particular, calcineurin subunits, CNI polypeptides, immunophilins and the like may be of mammalian origin, e.g., human origin.

### 25 VII. UTILITY

Methods and compositions of the present invention may be applied in a number of different ways. Following the guidance presented herein, one of skill in the art may isolate nucleic acids encoding additional CNI polypeptides, for example, a human CNI polypeptide.

In one approach, a yeast strain carrying a mutation of the CNI gene, e.g., a deletion, is used to clone heterologous sequences (e.g., human sequences) by complementation. A library of genomic DNA or, preferably, cDNA from an organism (e.g., human) and tissue (e.g., lymphocyte cells) of choice is cloned into a vector that can be maintained in yeast. Preferably, the vector contains a yeast promoter effective to express the heterologous sequences in yeast cells. Several heterologous libraries suitable for expression in Saccharomyces

cerevisiae containing DNA from S. pombe (Beach, et al.) and Drosophila have been constructed.

The library is transformed into a suitable yeast strain carrying a cni mutation, and transformants are selected using a suitable complementation assay. For example, transformants may be screened for increased hygromycin sensitivity, as experiments described herein indicate that cni deletion mutants possess a decreased sensitivity to hygromycin B (Example 8). The screen may be made more effective by using a yeast strain that is hypersensitive to hygromycin B, such as a strain deficient for a subunit of calcineurin (Example 8).

Alternatively, human CNI DNA sequences may be isolated by directly screening a library, e.g., a lymphocyte cDNA library, for clones hybridizing with a yeast CNI nucleic acid probe. The generation of an exemplary yeast CNI nucleic acid probe is described in Example 1.

In another approach, particularly advantageous for isolating sequences expressed at low levels, a CNI nucleic acid probe may be used to screen a genomic library, e.g., a human genomic library, to isolate a sequence that may be used to design probes or primers that may match the target sequence better that the yeast sequence. Such primers may be used with, for example, PCR, to isolate longer fragments from a tissue-specific library.

In yet another approach, an antibody generated against CNI polypeptide is used to immunoprecipitate a CNI polypeptide from an organism and/or tissue of choice. The protein may then be micro-sequenced, and the sequence utilized to design degenerate primers useful for isolating a cDNA.

CNI polypeptides of the present invention, particularly CNI fragments that retain a desired binding activity, may be used as lead compounds useful for the development of small molecules having cellular functions similar to those of the CNI-polypeptides, that is, molecules effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin.

CNI-polypeptides of the present invention may also be employed in a method of increasing sensitivity of cells to calcineurin-affecting immunosuppressant drugs. In this method, a CNI-polypeptide is introduced into the cell typically prior to or at the same time as contacting the cell with an immunosuppressant drug, such as FK506. The polypeptide may be delivered by any suitable means effective to deliver polypeptides to selected cells,

Alternatively, nucleic acids encoding CNI polypeptides may be used in appropriate expression vectors as a genetic therapy tool to potentiate the immunosuppressive effects of

calcineurin-targeting immunosuppressant drugs. The vectors may be targeted to selected cells. such as T-cells, to increase their sensitivity to a given systemic dose of an immunosuppressant.

Another utility of the present invention includes methods of screening for substances that up-regulate expression of CNI polypeptides, i.e., substances that affect transcription. Such 5 substances are useful for sensitizing cells to immunosuppressant drugs. In this method, the CNI promoter can be attached to a gene that functions as a selectable marker (for use in genetic selections to screen test substances) or to a reporter gene (for use in evaluating the effect on CMI transcription by test substances).

In another aspect of the present invention, the CNI-polypeptides, for example, mammalian homologue polypeptides of CNI, have potential use as therapeutic agents for both human and veterinary use. For example, CNI-polypeptides may be used in a method of enhancing immunosuppression in a test subject. In this method, the CNI-polypeptide is administered to the subject in a pharmaceutically-acceptable formulation and at a concentration effective to potentiate the interaction of an immunosuppressant/immunophilin complex with a subunit of calcineurin. The method may also include contacting the CNI-polypeptide with a cell under conditions effective to permit uptake of the protein into the cell in order to increase sensitivity of the cell to immunosuppressants. A CNI polypeptide used in such methods may be modified to be more suitable for administration or to be more effective in a cell. For example, a CNI polypeptide may be modified to eliminate PEST motifs, which are typically 20 found in proteins with short half-lives, to extend the effective lifetime of the polypeptide in the target cell.

The following examples illustrate, but in no way are intended to limit the present invention.

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### MATERIALS AND METHODS

Unless indicated otherwise, chemicals and reagents were obtained from Sigma Chemical Company (St. Louis, MO) or Mallinckrodt Specialty Chemicals (Chesterfield, MO), restriction endonucleases were obtained from New England Biolabs (Beverly, MA), and other modifying enzymes and biochemicals were obtained from Pharmacia Biotech (Piscataway, NJ) or Boehringer Mannheim (Indianapolis, IN). FK506 was obtained from Fujisawa USA, Inc. (Deerfield, IL), cyclosporin A was obtained from Sandoz (Basel, Switzerland), and rapamycin was obtained from Wyeth-Ayerst (Princeton, NI). Materials for media for yeast growth and culture were obtained from DIFCO (Detroit, MI). Unless otherwise indicated, manipulations

of yeast, bacteria, nucleic acids, proteins and antibodies were performed using standard methods and protocols (e.g., Guthrie, et al., Sambrook, et al., Ausubel, et al., Harlow, et al., and Rose, et al.).

#### 5 A. Buffers

Z buffer: 60 mM Na, HPO, -7H,O, 40 mM NaH, PO, -H,O, 10 mM KCl, 1 mM MgSO, - $7H_2O$  and 50 mM  $\beta$ -mercaptoethanol (pH 7.0).

#### B. Plasmids, Libraries and Yeast Strains

Plasmids pGBT9 (GBT), carrying GAL4 DNA-binding domain (amino acid residues 1-147; G4BD) and TRP1, and pGAD (GAD), carrying GAL4 activation domain (amino acid residues 768-881; G4AD) and LEU2; three pGAD libraries carrying fusions between G4AD and yeast genomic Sau3AI partial-digest fragments in each frame; and the yeast GAL1-locZ reporter strain SFY526 (Y526; MATa ura3-52 ade2-101 lys2-801 his3-200 trp1-901 leu2-3.112 15 can1 gal4-542 gal80-538 URA3::GAL1-lacZ) were obtained from Stanley Fields (State University of New York at Stony Brook, Stony Brook, NY; Chien, et al., Bartel, et al.). The libraries were constructed with linkers between the GAL4 activation domain and the Sau3Al fragments. The sequences of the linkers were 5'-ATCG-3' for the first library, 5'-ATCCG-3'for the second library, and 5'-ATCCCG-3'for the third library. In this way, the yeast genomic Sau3AI fragments were cloned in all three reading frames relative to G4AD.

Plasmids pAS2 (AS) carrying G4BD and TRP1, and pAS-lamin (AS-lamin) containing a sequence encoding a G4BD-lamin C fusion; and yeast reporter strains Y190 (MATa ura3-52 ade2-101 his3- $\Delta$ 200 trp1-901 leu2-3,112 cyh2 $\Delta$ , gal4\( \Delta \) gal80\( \Delta \) URA3::GAL-lacZ LYS2::GAL-HIS3), a derivative of Y153 carrying dual indicator genes (GAL-lacZ and GAL-25 HIS3), and Y187 (MATα ura3-52 ade2-101 lys2-801 his3-200 trp1-901 leu2-3,112 gal4Δ gal80\triangle URA3::GAL-lacZ) carrying GAL-lacZ reporter were obtained from Stephen Elledge (Baylor College of Medicine, Houston, TX; Durfee, et al.). Yeast strain Y153b1 (cnb1::ADE2) was derived from Y153.

E coli strain JBe181 (leuB600 trpC9830) was obtained from Ira Herskowitz (University 30 of California at San Francisco, San Francisco, CA). Protease-deficient yeast strain BJ2407 (Guthrie, et al.) was obtained from the Yeast Genetic Center (University of California at Berkeley, Berkeley, CA).

### C. GAL4-Calcineurin Fusions

GAL4-calcineurin (GAL4-CN) fusions, GBT-A1 (G4BD-CNA1), GBT-A2 (G4BD-CNA2), GBT-B1 (G4BD-CNB1), GAD-A1 (G4AD-CNA1), GAD-A2 (G4AD-CNA2), and GAD-B1 (G4AD-CNB1) were constructed as follows. Plasmids containing inserts encoding CN subunits CNA1 (SEQ ID NO:9; Cyert, et al., 1991), CNA2 (SEQ ID NO:11; Cyert, et al., 1991) and CNB1 (SEQ ID NO:13; Cyert, et al., 1992) were subjected to site-directed mutagenesis (Kunkle) to introduce a BamHI site just upstream of each subunit's initiation codon in the second reading frame. DNA prepared from the mutated plasmids was digested with BamHI and XhoI, and the resulting BamHI-XhoI fragments, each containing a full-length coding sequence, were cloned into GBT or GAD that had been cut with BamHI and Sal 1. The resulting plasmids encoded in-frame fusions of the CN subunits with G4BD or G4AD.

Plasmids encoding CNA protein variants with truncated C-termini (GBT-A1ΔC, GBT-A2ΔC) were constructed by introducing stop codons after amino acid residues 509 (CNA1) and 502 (CNA2). The 44-residue deletion in GBT-A1ΔC removed the autoinhibitory domain of CNA1, while the 102-residue deletion in GBT-A2ΔC removed both the autoinhibitory and the calmodulin-binding domains of CNA2 (Cyert, et al., 1991).

Plasmid GBT-FKBP, containing an FK506 binding protein (FKBP) gene fused to the GAL4 binding domain, was constructed by introducing a Bg/II site upstream of the initiation codon and a BamHI site downstream of the stop codon of FKBP12 (Heitman, et al.) and ligating the Bg/II-BamHI fragment into GBT cut with BamHI.

Plasmid B1/YEp352 was constructed to contain the full coding sequence of CNB1 (SEQ ID NO:13, Cyert, et al., 1992) as a 1.4 kb BamHI-EcoRI fragment encompassing the sequence presented as SEQ ID NO:13 (812 bp; contains the coding sequence), in the multicopy plasmid YEp352(HIS), which is derived from YEp352 (URA) (Hill, et al.).

Plasmid CNI/YEp352(HIS) (also referred to as CNIH) was constructed by ligating a 3.16 kb Munl-EcoRV fragment, containing the full coding sequence of CNI, from plasmid CNI7.1 (construction described below) into YEp352(HIS) cut with EcoRI and Smal. Plasmids CNI/YEp352(TRP) (also referred to as CNIT) and CNI/YEp352(URA) (also referred to as CNIU) were similarly constructed using the 3.16 kb fragment and YEp352(TRP) or YEp352(URA), respectively (Hill, et al.).

Plasmids A1/YEp351 and A2/YEp352 were constructed to contain the full coding sequences of CNA1 (SEQ ID NO:9, Cyert, et al., 1991) in YEp351 (Hill, et al.) and CNA2 (SEQ ID NO:11, Cyert, et al., 1991) in YEp351 and YEp352, respectively. A1/YEp351 was constructed by ligating a 2.9 kb Sacl-HindIII fragment from clone CNA1 (Cyert, et al., 1991)

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into YEp351(HIS) cut with Sacl and HindIII. A2/YEp352 was similarly constructed by ligating a 3 kb Spel-HindIII fragment from clone CNA2 (Cyert, et al., 1991) into YEp352(HIS) cut with Xbal and HindIII.

All GALA-CN fusions were verified by DNA sequencing (Sanger, et al.) using "SEQUENASE 2.0" sequencing kits (United States Biochemical, Cleveland, OH), and were subjected to the following tests. The functionality of the fusion proteins was assayed by determining whether they could complement the appropriate cn mutant phenotypes, using assays to measure the sensitivity to pheromone and Mn<sup>2+</sup> (Reneke, et al., Cyert, et al., 1991). All of the GAL4-CN fusions were functional in this assay.

The fusion proteins were also tested for their ability to activate the reporter gene in the absence of the complementary GAL-4 domain fusion (i.e., in the presence of the complementary GAL4 domain not fused to a second protein, for example, G4BD-A1 vs G4AD) using the two-hybrid interaction assay described below. Only GBT-B1 and GAD-A1 were able to activate the reporter gene at low levels without the complimentary GAL-4 domain fusion — assays with the other fusion proteins in the absence of the complimentary GAL-4 domain fusion showed no detectable levels of expression.

The two-hybrid interaction assay was also used to test the ability of the fusions to interact specifically with another fusion containing complimentary GAL4 and CN domains (e.g., G4BD-A1 interacting with G4AD-B1). All CN hybrids were able to react specifically and result in an activation of the reporter gene that was clearly detectable above background. The high specificity witnessed in these experiments indicates that the GAL4 two-hybrid system can reliably be used to assay interactions between CN and other proteins.

# D. Yeast GAL4 Two-Hybrid System for Detecting Protein-Protein Interaction

In the library screen, described in more detail in Example 1A, the yeast strain Y190, harboring the hybrid plasmid carrying the GAL4 binding domain fused to the A1 subunit of calcineurin (G4BD-CNA1), was transformed with fusion libraries carrying yeast genomic DNA Sau3Al fragments fused to the GAL4 activation domain. Transformants that were able to express the reporter genes, i.e., able to grow on -His + 3-AT and to score blue in  $\beta$ -gal assay, were selected as candidate positives. These candidate positives potentially contain library DNA fragments encoding proteins that physically interact with CNA1.

In another application described herein, the two-hybrid system was used to test for interactions between CNA (fused to one of the GAL4 domains) and CNB1 (fused to the other GAL4 domain), and between CNA and FKBP. Additional experiments tested a clone, CN1c,

isolated using the library screen, against a series of proteins fused to the complementary GAL4 domain under various conditions to test whether CN1c interacts with CNA subunits, and if so, how the interactions are affected by various conditions.

# E. Color Development (β-gal) Assay

Yeast reporters harboring both G4BD and G4AD fusions (and a third non-fusion plasmid in some cases) were monitored for  $\beta$ -gal activity as follows. Purified yeast transformants were patched onto selective plates with or without other test reagents. After growing 3 days at 30°C, colonies were lifted onto nitrocellulose filters, permeabilized in liquid nitrogen as above, placed on Whatman No. 1 paper in petri dishes containing 0.1% X-Gal in Z buffer (see above), and incubated at 30°C for 12 hours. Blue color begins to appear in positive colonies between about one half and ten hours into the incubation period.

Exemplary images obtained using the color development assay are presented in Figures 2A, 3A, 4A, 4C, 5A, 5C, 6A, 6C, 6F, 7A, 7C, 7E, 7G, 8A, 9A and 9C.

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# F. Growth Assay

A growth assay, applicable to yeast strains Y190 and Y153b1 which carry both GAL-HIS3 and GAL-lacZ reporters, was sometimes used as a complement to the color assay described above. Yeast transformants were streaked onto selective plates containing 40-50 mM 3-AT and no Histidine, and incubated at 30°C for 3-7 days. Growth (corresponding to the level of HIS3 expression) was monitored as an indicator of the interaction between fusion proteins. In cases where both assays were used, the amount of cell growth typically correlated well with the color intensity in the  $\beta$ -gal assay.

# 25 G. Yeast Growth, Drug Treatment

Yeast were typically grown in YPD (rich non-selective) or synthetic complete (SC) medium with selected component drop-outs, depending on the plasmid introduced, following standard procedures (Sherman, et al., Ausubel, et al.).

Experiments utilizing treatment with drugs or additives were performed by including the drug or additive in the medium. For plating, the agar was autoclaved, allowed to cool to  $50^{\circ}$ C, and the drug or additive was added before pouring the plates. Unless otherwise indicated, drugs and additives were added to result in the following final medium concentrations: FK506: 1  $\mu$ g/ml, cyclosporin A: 10  $\mu$ g/ml, rapamycin: 10 ng/ml, and hygromycin B:  $40 \mu$ g/ml.

### H. Antibodies

Polyclonal and monoclonal antibodies, for use in the present invention, can be prepared by standard methods (Harlow, et al.) utilizing the CNI polypeptides of the present invention, for example, a substantially purified CNI/β-galactosidase fusion protein (Example 9). Antibodies can also be generated by recombinant techniques (Cabilly, et al.; Better, et al.; Skerra, et al.). In addition to whole antibody molecules, antibody fragments retaining the immunological specificity of the whole antibody may also be used in the practice of the present invention (e.g., Fab and F(ab')<sub>2</sub> fragments of lgG (Pierce Chemical, Rockford, IL)). The antibodies can be purified by standard methods to provide antibody preparations which are substantially free of serum proteins that may affect reactivity (e.g., affinity purification (Harlow et al.)).

### EXAMPLE 1

### Isolation of CNIc

# A. <u>Library Screening</u>

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Yeast strain Y190 was transformed with pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid using the transformation protocol described by Schiestl, et al. Transformants were selected, colony purified, and a single transformant was selected to make (Y190 GBT-A1)-competent cells, following the procedure described in Guthrie, et al..

The three pGAD yeast fusion libraries described above, carrying fusions between G4AD and yeast genomic DNA Sau3Al fragments in each reading frame, were then used to transform (Schiestl, et al.) the Y190 GBT-A1-containing cells. Transformants were plated onto SC-Trp-Leu-His plates containing 40 mM 3-aminotriazole (3-AT; Sigma Chemical Co., St. Louis, MO) and incubated at 30°C for 6 days to screen for HIS colonies (Durfee, et al.).

His \* colonies were replica plated onto nitrocellulose filters (Schleicher & Schuell, Keene, NH), frozen in liquid nitrogen for approximately 30 seconds, and incubated at 30 °C for 12 hours with Z buffer (see above) containing the chromogenic substrate X-Gal (0.1%) to assay  $\beta$ -gal activity (Breeden, et al.).

Candidate positive (blue) colonies were re-streaked for single colonies. Single colonies were purified and retested using the above protocol. Colonies which reproducibly tested positive were screened using PCR with primers directed against the internal portion of GAL-4 (i.e. the portion between the DNA binding domain and the activation domain). The sequences of the primers, G4-PCR-A and G4-PCR-B, are given as SEQ ID NO:22 and SEQ ID NO:23,

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respectively. Colonies yielding a PCR product were identified as containing intact GAL4, and were eliminated.

The GBT-A1 TRP\* plasmid was eliminated by growing in Trp\* liquid media for 2-3 days, plating on -Leu media and then replica-plating on -Leu and -Trp plates to identify and eliminate colonies that had lost the GBT-A1 plasmid, yet still gave a positive signal.

Plasmid DNA was extracted from the remaining Leu\* candidates. The plasmid DNA was transformed into E coli JBe181 and plated on -Leu media to select for library plasmids. The library plasmids isolated by this method were introduced back to the yeast reporter strains either alone or with test G4BD fusions: GBT, GBT-A1, and AS-lamin.

A parallel specificity assay was conducted by mating. Candidate strains, as described above, were 3-AT growth positive and X-gal positive when both the library and GBT-A1 plasmids were present. After elimination of the GBT-A1 plasmid from these strains, strains that were Leu Trp 3-AT growth and  $\beta$ -gal were mated to the following strains: Y187 (MATa) carrying GBT, GBT-A1, or AS-lamin, and the diploids were assayed.

Among the 3-AT positive,  $\beta$ -gal positive candidates identified by the secondary screening method just described, one clone (III-21S, later termed GAD-CNIc) was specifically positive in conjunction with GBT-A1 in both the transformation assay and the mating assay.

# B. Sequence of CNIc

Clone III-21S was sequenced as above. The sequence is presented herein as SEQ ID NO:1, and a schematic representation of the clone is shown in Figure 1A. The Sau3AI library insert encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GAL4, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product.

25 Accordingly, the clone was termed CNIc, with the lowercase "c" representing "c-terminal".

The stippled portion between GAL-4AD and CNIc in Figure 1A represents the linker discussed in Materials and Methods, above. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons, and Sau3AI restriction sites.

# 30 C. Isolation of a Full Length Clone

A <sup>32</sup>P-labeled CNIc probe was generated from the 1.22 kb CNIc insert of clone III-21S by polymerase chain reaction (PCR) using primers represented as SEQ ID NO:7 and SEQ ID NO:8. The probe was used to map the gene to the right arm of chromosome 11 by hybridization screening (Sambrook, *et al.*) a panel of  $\lambda$  clones (American Type Culture

Collection (ATCC), Rockville, MD) spanning the entire yeast genome. Two clones, 70500 and 70590, gave positive hybridization signals. A phage lysate of clone 70500 in \( \lambda \) MG3 was obtained from the ATCC, was amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments.

The phage DNA was digested with SacI, yielding a 7.1 kb fragment containing the entire CNI gene. This fragment was cloned into "BLUESCRIPT SK" (Stratagene, La Jolla CA) cut with Sacl, yielding plasmid CNI7.1. Plasmid CNI7.1 was digested with Munl and EcoRV, releasing a 3.16 kb fragment containing the entire coding sequence of CNI. The 3.16 kb fragment was then cloned into each of YEp352(HIS), YEp352(TRP), YEp352(URA), and 10 "BLUESCRIPT SK", each cut with EcoRl and Smal, yielding plasmids CNIH, CNIT, CNIU and CNI3.2, respectively. The sequence of the 3.16 kb MunI/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4. The Munl site is at nucleotide 100 of SEQ ID NO:4, and the EcoRV site is at nucleotide 3263 of SEQ ID NO:4.

A schematic diagram of the sequence presented as SEQ ID NO:4 is shown in Figure 1B. This sequence contains the 3.16 kb Munl/EcoRV fragment used in many of the experiments described herein (depicted in Figure 1B as the portion between the Mun1 and EcoRV sites), which contains the entire 2.75 kb coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

Figure 13 also shows the location of certain features of the sequence. For example, "PEST" motifs (Rogers, et al., Dice) are indicated by bars over the corresponding sequence.

A search of known DNA and protein sequences turned up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin-binding protein.

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### EXAMPLE 2

# Binding of CNIc to Calcineurin

Y190 yeast carrying the plasmids indicated below were assayed for  $\beta$ -gal activity by color development assay described above to determine the specificity of binding of CNIc to subunits of calcineurin.

Exemplary data, in the form of images of filters having yeast colony replicas that had undergone the  $\beta$ -gal color development assay are shown in Figure 2A. The legend for Fig. 2A is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs. The constructs are as follows: 20:GBT-A1 and

GAD-B1, 21:GBT-A1 and GAD-CNIc, 22:GBT-A1 and GAD-CNIc, 23:GBT-A1 and GAD-CNIc 24:GBT-A2 and GAD-CNIc 25:AS-lamin and GAD-CNIc 26:GBT and GAD-CNIc.

Yeast colonies used in the assay were derived by several different methods. Those at location 22 were purified colonies from the original library screen, those at 21 were colonies transformed with mini-prep DNA of the isolated GAD-CNIc plasmid, and the remaining colonies (23, 24, 25 and 26) were transformed with maxi-prep (Qiagen, Chatsworth, CA) DNA of GAD-CNIc.

A comparison of the intensities of the blue β-gal reaction product indicates that CNIc interacted strongly with CNA1 regardless of the source of the CNIc plasmid DNA (20, 21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with GAD-CNIc (26) showed a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact were used as a positive control for the assay (20).

In summary, the data above show that CNIc interacted specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments was conducted using constitutively-active CNA subunits, as well as calcineurin subunit CNB1. As described in Materials and Methods, above, CNA1ΔC and CNA2ΔC were each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A. The legend for Fig. 3A is shown in Figure 3B. Locations of yeast colonies expressing specific constructs are as follows: 27:GBT and GAD-CNIc, 28:GBT-A1ΔC and GAD-CNIc, 29:GBT-A2ΔC and GAD-CNIc, 30:GBT-A1ΔC and GAD, 31:GBT-A2ΔC and GAD, 32:GBT-B1 and GAD-CNIc, 33:GBT-B1 and GAD, 34:GBT-A1ΔC and GAD-B1, and 35:GBT-A2ΔC and GAD-B1.

The data show that GBT-A1 $\Delta$ C and GAD-CNIc (28) gave a definite positive signal, while GBT-A2 $\Delta$ C and GAD-CNIc (29) was weaker, though still detectable above its background (i.e. GBT-A2 $\Delta$ C and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) was not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 $\Delta$ C and GAD-B1 (34) and GBT-A2 $\Delta$ C and GAD-B1 (35) gave strong signals.

These data show that CNIc interacted specifically with CNA1ΔC and CNA2ΔC, but not with CNB1.

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### EXAMPLE 3

# Effects of Immunosuppressant Drugs on Binding of CNIc to Calcineurin in B1. B1 Deletion and B1 Overproducing Yeast Strains

Three yeast strains carrying the plasmids indicated below were assayed for  $\beta$ -gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments were performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data, in the form of filter images produced as above are shown in Figures 4A, 4C, 5A and 5C. Plates used to make the filters shown in Figs. 4A and 4C were replicas from one master plate, while plates used to make the filters shown in Figs. 5A and 5C were replicas from another plate. The plates used to generate filters shown in Figs. 4A and 5A were without FK506, while the plates used to generate filters shown in Figs. 4C and 5C contained  $1 \mu g/ml$  FK506.

The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Since the imaged filters in Figs. 4A and 4C were from replica plates, corresponding locations on each filter contain material from the same yeast colonies. Accordingly, the locations are referred to by the same "base" numbers in the legends. To facilitate reference to a specific location on a specific filter, the base numbers are followed by a lowercase letter that is different for each of the individual filters. For example, in the present figure, "a" follows the base numbers to identify locations on the filter shown in Fig. 4A, while a "b" follows the base numbers to identify locations on the plate in Fig. 4C. This labeling scheme is used in other experiments detailed herein where multiple filter lifts are shown.

The interactions of various combinations of proteins expressed by constructs indicated below was studied in three yeast strains. Strain Y153b, at 36-40, is null for the CNB1 subunit of calcineurin. Strains Y190 (41-46) and Y526 (47-51) are wild-type for CNA1, CNA2 and CNB1.

Hybrid proteins expressed by colonies at specific locations are as follows: 36:GBT-A1 and GAD-CNIc, 37:GBT-A2 and GAD-CNIc, 38:GBT-A1ΔC and GAD-CNIc, 39:GBT and GAD-CNIc, 40:GBT-A2ΔC and GAD-CNIc, 41:GBT-A1 and GAD-CNIc, 42:GBT and GAD-CNIc, 43:GBT-A2 and GAD-CNIc, 44:GBT-A1ΔC and GAD-CNIc, 45:GBT-A1 and GAD-B1, 46:GBT-A2ΔC and GAD-CNIc, 47:GBT-A1 and GAD-CNIc, 48:GBT and GAD-CNIc,

49:GBT-A2 and GAD-CNIc, 50:GBT-A1ΔC and GAD-CNIc, and 51:GBT-A2ΔC and GAD-CNIc.

Yeast strain Y526 was used for all experiments shown in Figs. 5A, 5B, 5C and 5D. The expression vector B1/YEp352(HIS) was not used in strains Y190 or Y153b1 because they are HIS\* in the absence of 3-AT.

The base numbers in Figures 5B and 5D correspond to locations of colonies expressing the following constructs: 52:GBT-A1, GAD-CNIc and YEp352, 53:GBT-A1, GAD-CNIc and B1/YEp352, 54:GBT-A2, GAD-CNIc and YEp352, 55:GBT-A2, GAD-CNIc and B1/YEp352, 56:GBT, GAD-CNIc and B1/YEp352, 57:GBT-A1ΔC, GAD-CNIc and B1/YEp352, and 58:GBT-A2ΔC, GAD-CNIc and B1/YEp352.

A comparison of data shown in Fig. 4A (no added drugs) shows the effect of deleting the endogenous host CNB1 gene on interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1ΔC and CNA2ΔC. Note that interactions in panels 36a-40a (CNB1 null strain) were all stronger (with the exception of the negative control in 39) than interactions in corresponding panels 41a-51a (strains wild-type for CNB1). This result indicates that interaction between CNIc and CNA subunits were enhanced by the deletion of the CNB1 subunit.

Comparison of corresponding panels in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA\(Delta\)C interactions. The drug enhanced interactions under all except control (39, 42 and 48) conditions. The effect was most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhanced CNIc-CNA/CNA\(Delta\)C interactions under conditions where the CNB1 subunit was overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNA\(Delta\)C interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) had reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhanced interactions in all colonies, except the negative controls (56).

Taken together, the above data demonstrate that the interaction of CNIc with CNA and CNAΔC was markedly enhanced by FK506. The interaction was also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction was overcome by the stimulatory effect of FK506.

In light of the effects of FK506 on CNIc-CNA/CNA $\Delta$ C interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H. Filters shown in

Figs. 6A-6F were from replica plates, as were those in Figs. 7A-7H. Colonies shown in Figs. 6A and 7A were plated without drugs; those in Figs. 6C and 7C were plated with FK506 (1 µg/ml), those in Figs. 6E and 7E with CsA (10 µg/ml), and those in Fig. 7G with rapamycin (10 ng/ml). Yeast strains used were as follows: In Figs. 6A-6F, panels 59-63 were Y153b1; 5 64-69 were Y190, and 70-74 were Y526. In Figs. 7A-7H, panels 77-83 were Y526, and panels 75 and 76 were Y190.

The base numbers in Figures 6B, 6D and 6F correspond to locations of colonies expressing the following constructs: 59:GBT-A1 and GAD-CNIc, 60:GBT-A2 and GAD-CNIc, 61:GBT-A1ΔC and GAD-CNIc, 62:GBT and GAD-CNIc, 63:GBT-A2ΔC and GAD-CNIc, 10 64:GBT-A1 and GAD-CNIc, 65:GBT and GAD-CNIc, 66:GBT-A2 and GAD-CNIc, 67:GBT-A1AC and GAD-CNIc, 68:GBT-A1 and GAD-B1, 69:GBT-A2AC and GAD-CNIc, 70:GBT-A1 and GAD-CNIc, 71:GBT and GAD-CNIc, 72:GBT-A2 and GAD-CNIc, 73:GBT-A1AC and GAD-CNIc, and 74:GBT-A2\Delta and GAD-CNIc.

The base numbers in Figures 7B, 7D, 7F and 7H correspond to locations of colonies expressing the following constructs: 75:GBT-FKBP and GAD, 76:GBT-FKBP and GAD-CNlc. 77:GBT-A1, GAD-CNIc and YEp352, 78:GBT-A1, GAD-CNIc and B1/YEp352, 79:GBT-A2, GAD-CNIc and YEp352, 80:GBT-A2, GAD-CNIc and B1/YEp352, 81:GBT, GAD-CNIc and B1/YEp352, 82:GBT-A1ΔC, GAD-CNIc and B1/YEp352, and 83:GBT-A2ΔC, GAD-CNIc and B1/YEp352.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. The constructs and yeast strains at corresponding locations were the same. As expected, the  $\beta$ -gal signal was also essentially equivalent between the two sets. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A was also effective in enhancing interaction of CNIc with CNA and CNAAC. 25 Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through inhibition of calcineurin activity (Cyert).

Similarly, data shown in Figures 7A and 7C are essentially equivalent to those in Figures 5A and 5C, except that a top panel has been added in Figs. 7A-H. As above, the corresponding panels show the same constructs and yeast strains. The added panels (75 and 76) assessed the interaction of an FK506 binding protein (FKBP) with CNIc, and indicate that there were no detectable interactions between these proteins. Results in Fig. 7E demonstrate that cyclosporin A had a similar effect to FK506 in cells overexpressing CNB1 -- that is, it enhanced the interactions between CNIc and CNA/CNAAC.

In contrast, data presented in Fig. 7G show that the immunosuppressant rapamycin, which is known not to target calcineurin, had no detectable effect on CNIc-CNA/CNA $\Delta$ C interactions (compare Fig. 7G with Fig. 7A).

Taken together, the above data show that like FK506, cyclosporin A (CsA), but not rapamycin, also enhanced the interaction of CNIc with CNA and CNAΔC. CNIc didn't interact with FKBP with or without FK506.

### EXAMPLE 4

### Effects of CNI on FKBP/FK506 binding to Calcineurin

Y526 cells, carrying the plasmids indicated below, were grown in -Trp-Leu-His liquid media with or without FK506 (1 μg/ml) until OD<sub>600</sub> reached about 1.0. Approximately the same number of cells, calculated based on OD<sub>600</sub> and equivalent to 1 ml of an OD<sub>600</sub>=1 suspension, was harvested from each culture, washed once with ddH<sub>2</sub>O, centrifuged briefly, and the pellet was resuspended in 30 μl ddH<sub>2</sub>O and transferred onto a nitrocellulose filter. The filters were frozen in liquid nitrogen as described above, placed in a 8.5 cm petri dish containing a sheet of Whatman No. 1 paper (Whatman International LTD, Maidstone, UK) in 1.6 ml Z buffer containing 0.1% X-Gal, and incubated at 30°C for 8 hours.

Figure 8A presents exemplary data from studies to assess the effect of CNI overexpression on FK506-mediated FKBP interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Locations of yeast colonies expressing specific constructs: 84:GBT-FKBP, GAD-A2 and YEp352, 85:GBT-FKBP, GAD-A2 and YEp352, 86:GBT-FKBP, GAD-A2 and CNI/YEp352, and 87:GBT-FKBP, GAD-A2 and CNI/YEp352. The cells at 85 and 87 were exposed to FK506, while those at 84 and 86 were not.

Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 showed no detectable interaction (84). In the presence of FK506, however, the proteins interacted (85), presumably because FK506 formed a complex with FKBP, which then bound CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI had no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiated, or enhanced the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

Experiments illustrated in Figures 9A and 9B demonstrate that CNI overproduction had little or no effect on the binding of CNA2 to CNB1, providing support for the specificity of the stimulatory effect that CNI overproduction had on the FK506-dependent binding of FKBP to calcineurin. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively.

Locations of yeast colonies expressing the following constructs: 88:GBT-A2, GAD-B1 and YEp352, 89:GBT-A2, GAD-B1 and YEp352, 90:GBT-A2, GAD-B1 and CNI/YEp352, and 91:GBT-A2, GAD-B1 and CNI/YEp352. The colonies at 89 and 91 were exposed to FK506, while colonies at 88 and 90 were not.

Taken together, the above data show that overexpression of the full-length CNI clone markedly enhanced the FK506-dependent interaction of FKBP with CNA, although it didn't affect the interaction between CNA and CNB1.

### EXAMPLE 5

## Co-Immunoprecipitation of CNIc and CNA

Yeast BJ2407 harboring AS-CNIc, which carries an influenza hemagglutinin (HA) epitope tag (Wilson, et al.), and GAD-A2 (lanes 1, 3) or A2/YEp352 (lanes 2, 5), and strain MCY300-1 (cna1 cna2; lane 4) were grown in selective media to  $OD_{eco}=0.8$ . The cells were harvested, lysed, and immunoprecipitated in the presence of 25  $\mu$ g/ml FK506 with anti-HA monoclonal antibody (obtained from M. Kirschner, Harvard University, Boston, MA; Wilson, et al.), following protocols described in Harlow, et al. The cell extracts (lanes 3-5) and the immune complex (lanes 1, 2) were resolved by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; Laemmli) followed by western blot with a rabbit anti-CNA2 polyclonal antibody generated using standard methods (Harlow, et al.). Bound anti-CNA2 antibody was visualized with the "ECL" kit (Amersham, Arlington Heights, IL) using goat antirabbit antibody. Molecular weight markers are indicated on the right in kD.

The results demonstrate that CNIc was capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. This independent, biochemical assay confirmed the results described above obtained using the two hybrid protein interaction assay — that is, that CNIc physically interacted with and bound CNA subunits.

Cell extracts of BJ2407 harboring AS-CNIc, and GAD-A2 or A1/Yep351, and Y153b1 harboring AS-CNIc and GAD-A1 were subjected to SDS-PAGE followed by western blot with anti HA antibody. The results showed that CNIc was present at very low levels in vivo.

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The results are consistent with the observations that a limited amount of CNA2 was precipitated by anti-HA antibody recognizing the CNIc fusions, and that CNI contains PEST-like motifs, a feature of proteins with a short half-life in vivo (Rogers, et al.).

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# EXAMPLE 6

## Northern Blot of CNIc

Norther blots (e.g., Sambrook, et al.) of yeast total RNA were hybridized with a CNIc probe. Exemplary data are shown in Figure 11. 20  $\mu$ g yeast RNA from YPH499 (lane 1) and MCY300-1 (lane 2) was resolved in a formaldehyde-agarose gel, transferred onto "HYBOND N\*" membrane (Amersham, Arlington Heights, IL), and hybridized with  $5x10^{\circ}$  cpm/ml probe of the CNIc insert (1.22kb). A single message of approximately 2.9 kb was detected in both strains at about the same level following an 18-hour exposure on XAR5 film (Eastman Kodak, Rochester, NY).

The data indicate that CNI was a physiologically expressed gene encoding a 2.9 kb message in yeast.

### EXAMPLE 7

# Chromosome Mapping of CNIc

A yeast chromosome blot obtained from the ATCC was hybridized with probe of the CNIc insert following the Southern hybridization procedure described in Sambrook, et al. A positive hybridization signal was obtained with two ATCC yeast genomic λ clones derived from chromosome 11. Clone 70500 had a relatively strong signal, while clone 70590 had a somewhat weaker one. A phage lysates of clone 70500 was ordered from the ATCC, amplified, purified, restriction-mapped, and used as a DNA source for cloning full length CNI (Example 1).

# EXAMPLE 8

# CNI null Mutants

## 1. Construction of cni Null Mutation

A 5', 1.8 kb Bg/II-HindIII and a 3', 0.9 kb Xbal-Bg/II fragment of CNI were ligated into pRS305(LEU2) (Sikorski, et al.). The resultant plasmid had a deletion of a 2 kb HindIII-Xbal fragment from the coding sequence of CNI. This cni::LEU2 mutant was introduced into the genomes of yeast haploid strains YPH499 (Sikorski, et al.), MCY300-1 (cna1 cna2) and DD12 (cnb1) (Cyert, et al., 1991, Cyert, et al., 1992) as well as two diploid strains.

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Leucine prototrophs were isolated at high frequency from all strains, and hybridization analysis confirmed that the cni::LEU2 allele had replaced the CNI gene. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains (even cni cn double mutants) can survive.

CNI was deleted from three yeast strains: YPH499 (WT), MCY300-1 (cna1 cna2), and DD12 (cnb1), resulting in cni strains LHy499, LHy300 and LHy12, respectively. Cells representing four colonies of each cni knockout strain and two colonies of each parent strain were grown in liquid YPD (Sherman, et al.) to saturation. Same numbers of cells from each culture were then plated onto YPD+Hygromycin B (40  $\mu$ g/ml) and growth was monitored at 30°C.

CNI deletions in each strain rendered that strain more resistant to hygromycin B. The effect was particularly pronounced in both MCY300-1 and DD12, suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B. The data indicate that deletion of CNI results in higher resistance to hygromycin B.

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## EXAMPLE 9

# Isolation of CNI/B-Galactosidase Fusion Protein

A CNI coding sequence is cloned into the  $\lambda$  gt11 vector (Stratagene, La Jolla, CA). The coding frame is cloned in-frame to the  $\beta$ -galactosidase coding sequences present in  $\lambda$  gt11. Bacterial lysogens infected either with lambda phage gt11 or with gt11/CNI are incubated in 32°C until the culture reaches to an O.D. of 0.4. Then the culture is incubated in a 43°C water bath for 15 minutes to induce gt11 peptide synthesis, and further incubated at 37°C for 1 hour. Bacterial cells are pelleted and lysed in lysis buffer (10 mM Tris, pH 7.4, 2 % "TRITON X-100" and 1% aprotinin). Bacterial lysates are clarified by centrifugation (10K, for 10 minutes, Sorvall JA20 rotor) and the clarified lysates are incubated with Sepharose 4B beads conjugated with anti- $\beta$ -galactosidase (Promega).

Binding and elution of  $\beta$ -galactosidase fusion proteins are performed according to the manufacturer's instruction. Typically binding of the proteins and washing of the column are done with lysis buffer. Bound proteins are eluted with 0.1 M carbonate/bicarbonate buffer, pH 10.

While the invention has been described with reference to specific methods and embodiments, it will be appreciated that various modifications and changes may be made without departing from the invention.

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: The Board of Trustees of the Leland Stanford Junior University
  - (ii) TITLE OF INVENTION: Calcineurin Interacting Protein Compositions and Methods
  - (iii) NUMBER OF SEQUENCES: 23
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: Dehlinger & Associates
    - (B) STREET: 350 Cambridge Avenue, Suite 250
    - (C) CITY: Palo Alto
    - (D) STATE: CA
    - (E) COUNTRY: USA
    - (F) 21P: 94306
  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
  - (vi) CURRENT APPLICATION DATA:

    - (A) APPLICATION NUMBER: (B) FILING DATE: 23-OCT-1995
    - (C) CLASSIFICATION:
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 08/328,322
    - (B) FILING DATE: 24-OCT-1994
  - (viii) ATTORNEY/AGENT INFORMATION:

    - (A) NAME: Sholtz, Charles K.
      (B) REGISTRATION NUMBER: P38,615
    - (C) REFERENCE/DOCKET NUMBER: 8600-0151.41
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: (415) 324-0880
      - (B) TELEFAX: (415) 324-0960
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1222 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (C) INDIVIDUAL ISOLATE: Sau3Al fragment containing CNIc coding sequence
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS (B) LOCATION: 1..918

(xi) SEQUENCE I	DESCRIPTION:	SEQ ID	NO:1:
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							Gln			ATT	48
						Leu		CCT Pro	Ala	TTA Leu	96
					Ile			CAA Gln 45		TCA Ser	144
								AAG Lys			192
				Pro				TAT Tyr			240
								ACA Thr			288
								ACA Thr			336
								GAT Asp 125			384
								TTC Phe			432
								CAC His			480
								AAA Lys			528
								AGA Arg			576
CAG Gln								AGT Ser 205			624
								AAA Lys			672
								AAT Asn			720
		λen						GTC Val			768

GCA TTT TAT GAT CCA AGA Ala Phe Tyr Amp Pro Arg 260			
GTA AGC AAG AAC CAT GTT Val Ser Lys Abn His Val 275			
AGT GAA AAC CGT GTA CTG Ser Glu Ann Arg Val Leu 290			
AGA AAA TAAGTACATT ATTT Arg Lys 305	CATTC TCCGACAGA	AN TIGCTACCAT TITACTITG	T 968
GTCCTGTGAT TCAATAGTGT A	CANTATATT GGACAT	TTTTA TAGTATACAA ATATAC	ACCA 1028
TCAATCTATA CATCCATATC A	CTTGTCGTA AAGATA	ATCCC TTTTTAATAG TACAGO	GATT 1088
AAAAAAATAA CATGATTAAC G	TTCAGTTAC CAATG	AGCTT ATTTATTAGG CTTGCT	TTAG 1148
ATTTTTCCAA GTCAATTTTT G	ITTTTCTA ACGCT	GCAA CCTCATCTCA ACCTTC	TTCC 1208
TTTGCAAGCA GATC			1222

## (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 306 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Asp Gln Ser Ser Asn Val Phe Ala Ser Lys Gln Leu Val Ala Asn Ile 1 5 10

Tyr Lys Pro Asn Gln Ile Pro Arg Glu Leu Thr Ser Pro Gln Ala Leu 20 25 30

Pro Leu Ser Pro Ile Thr Ser Pro Ile Leu Asn Tyr Gln Pro Leu Ser 35 40

Asn Ser Pro Pro Pro Asp Phe Asp Phe Asp Leu Ala Lys Arg Gly Ala 50 60

Ala Asp Ser His Ala Ile Pro Val Asp Pro Pro Ser Tyr Phe Asp Val 65 70 75 80

Leu Lys Ala Asp Gly Ile Glu Leu Pro Tyr Tyr Asp Thr Ser Ser Ser 90 95

Lys Ile Pro Glu Leu Lys Leu Asn Lys Ser Arg Glu Thr Leu Ala Ser 100 105 110

Ile Glu Glu Amp Ser Phe Amn Gly Trp Ser Gln Ile Amp Amp Leu Ser 115 120 125

Asp Glu Asp Asp Asn Asp Gly Asp Ile Als Ser Gly Phe Asn Phe Lys 130 135 140

Leu Ser Thr Ser Ala Pro Ser Glu Asn Val Asn Ser His Thr Pro Ile 145 150 155 160 WO 96/12806

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									••						
Leu	Gln	Ser	Leu	Aen 165	Met	Ser	Leu	Авр	Gly 170	Arg	Lys	Lys	Asn	Arg 175	Ala
Ser	Leu	His	Ala 180	Thr	Ser	Va]	Leu	Pro 185	Ser	Thr	Ile	λrg	Gln 190	λsn	Asn
Gln	His	Phe 195	λen	Asp	lle	Aen	Gln 200	Met	Leu	Gly	Ser	Ser 205	Asp	Glu	Amp
Ala	Phe 210	Pro	Lys	Ser	Gln	Ser 21!	Leu	Asn	Phe	λsn	Lys 220	Lys	Leu	Pro	Ile
Leu 225	Lys	Ile	Asn	Asp	Aen 230	Val	lle	Gln	Ser	Авл 235	Ser	Asn	Ser	Asn	Asn 240
Ъrg	Val	увь	yau	Pro 245	Glu	Хвр	Thr	Val	Asp 250	Ser	Ser	Val	Asp	11e 255	Thr
Ala	Phe	Tyr	<b>Asp</b> 260	Pro	Arg	Met	Ser	Ser 265	qax	Ser	Lys	Phe	Asp 270	Trp	Glu
Val	Ser	Lys 275	A∎n	Hie	Val	Asp	Pro 280	Ala	Ala	Tyr	Ser	Val 285	Asn	Val	Ala
Ser	Glu 290	Asn	Àrg	Val	Leu	Авр 295	Двр	Phe	Lys	Lys	Ala 300	Phe	λrg	Glu	Lys
Arg 305	Lys														
(2)	INFO	RMAT	TION	FOR	SEQ	ID N	0:3:								
	(i)	() (E	() LE () TY () ST	NGTH PE: RAND	nucl	TERI 8 ba eic SS: unkn	se p scid doub	airs							
	(ii)	HOL	.ECUL	E TY	PE:	CDNA	to	mRNA							
(	111)	нүр	OTHE	TICA	L: N	0									

- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: CINc coding sequence
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GATCAAAGTA GCAATGTCTT CGCATCCAAA CAGCTGGTCG CAAACATTTA TAAGCCCAAT 60 CAGATTCCAA GAGAATTAAC TTCTCCTCAG GCGTTACCAT TATCGCCCAT CACCTCACCA 120 ATTCTCAATT ACCAACCATT ATCAAACTCC CCGCCTCCAG ATTTTGATTT TGATCTAGCT 180 AAGCGCGCC CAGCCGATTC TCATGCTATT CCTGTGGATC CTCCATCATA TTTTGATGTA 240 TTANAGGCCG ATGGGATTGA ATTGCCATAC TACGATACAA GTTCATCTAA AATTCCTGAA 300 CTANANCTAN ACANATCTAG AGAGACATTG GCCAGCATTG AGGAGGACTC ATTCANTGGT 360 TGGTCTCAAA TTGATGACTT ATCCGACGAA GATGACAATG ATGCCGATAT AGCATCTGGT 420 TTCAACTTCA AGCTGTCAAC CAGTGCTCCG AGTGAGAACG TTAATTCACA CACTCCTATT 480 TTGCAGTCTT TANACATGAG TCTTGATGGG AGAAAAAAA ATCGTGCCAG TCTACACGCA 540

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ACATCAGTGT	TACCTAGTAC	AATAAGACAG	AACAATCAGC	ATTTCAATGA	CATAAACCAG	600
ATGCTAGGCA	GTAGTGACGA	AGATGCCTTT	CCCAAAAGCC	AATCATTAAA	TTTCAATAAG	660
AAACTACCAA	TACTTAAAAT	TAATGATAAC	GTCATACAAT	CAAACAGCAA	TAGTAATAAC	720
AGAGTTGATA	ATCCAGAAGA	TACAGTGGAT	TCTTCAGTCG	ATATTACAGC	ATTTTATGAT	780
CCAAGAATGT	CATCAGATTC	CAAATTTGAT	TGGGAGGTAA	GCAAGAACCA	TGTTGACCCA	840
GCAGCCTACT	CGGTTAACGT	TGCTAGTGAA	AACCGTGTAC	TGGACGACTT	TAAGAAAGCA	900
TTTCGCGAAA	AGAGAAAA					918

# (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3500 base pairs (B) TYPE: nucleic scid

  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNI coding sequence
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 376.,3120
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CCGAACACTT CCTTCGAGAG AGTGCATTTT ACTATGTGAA CCAATTTTTC CTCTTTTTCG	60
GTTTGCAAGT TCACCTGAAA AACTGCTTAA CACTACTAGC AATTGCCCTA TTGTCGTACG	120
AGGACTITGC CAAATGTATT CCCGGCTGTT TGTAGTATAT ATACGCAGAT ATATAATAGC	180
GCCGTCTTTT TACCTCTTTG AGCGAATTGC CAAATATTGA CTCTTTTGTC TTATTTCGCT	240
ATCCCCATCT TATCAAAAAT GGGAACAACT CGTTGAAATA AGAGACAAGC AACAAGAAAG	300
ACAACCAACA GAAAGTTCCA TTCCGCACAA ATACGCTGGA ATCCCATAGA ATATTGCTTG	360
TTCCTCTATG ACTAC ATG CTC CAA TTC AAT ACA GAA AAT GAT ACT GTA GCT Het Leu Gln Phe Amn Thr Glu Amn Amp Thr Val Ala 1 5 10	411
CCA GTG TTT CCC ATG GAG CAA GAT ATA AAT GCA GCA CCT GAT GCC GTC Pro Val Phe Pro Het Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val 15 20 25	459
CCA CTG GTG CAG ACA ACA ACA CTA CAA GTC TTT GTA AAG CTT GCC GAA Pro Leu Val Gln Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu 30 35 40	507
CCC ATA GTG TTT TTA AAA GGA TTT GAA ACT AAC GGA CTG TCT GAA ATA Pro lle Val Phe Leu Lym Gly Phe Glu Thr Amn Gly Leu Ser Glu Ile 45 50 60	555

															CCG Pro	603
															ACA Thr	651
															GTT Val	699
			GTC Val												GGC Gly	747
			TTC Phe												AAT Asn 140	795
			ATG Het													843
			ATC 11e 160													891
			GAT ABP													939
			CCA Pro													987
			CAA Gln													1035
			AAT Asn													1083
			TTC Phe 240													1131
CCT Pro	GAG Glu	CAA Gln 255	GA <b>G</b> Glu	GAT Asp	AAC Asn	TAT Tyr	CTT Leu 260	ACA Thr	CCA Pro	TCC	AAA Lys	GAT Asp 265	TCT Ser	YYY Lys	G <b>AA</b> Glu	1179
GTT Val	TTT Phe 270	ATT	TTT Phe	CGA	CCG Pro	GGC Gly 275	GAT Asp	TAT Tyr	ATT Ile	TAC Tyr	ACT Thr 280	TTT Phe	GAA Glu	CAG Gln	CCA Pro	1227
ATA 11e 285	TCG Ser	CAA Gln	TCT Ser	TAT Tyr	CCA Pro 290	GAA Glu	AGT Ser	ATA Ile	AAA Lys	GCC Ala 295	AAT Amn	TTT Phe	GGT Gly	TCC Ser	GTG Val 300	1275
			CTG Leu													1323
ACT Thr	ATA Ile	CAT His	ACT Thr 320	C <b>AA</b> Gln	TTA Leu	CCC Pro	ATC Ile	AAA Lys 325	GTC Val	GTA Val	λ <b>G</b> G λ <b>r</b> g	CTT Leu	CCT Pro 330	TCT Ser	GAT Abp	1371

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					GCA Ala				AAA Lys	1419
					TTC Phe		Glu			1467
	_				TTC Phe					1515
					ACA Thr 390					1563
					GCT Ala					1611
					GGT Gly					1659
					AAA Lys					1707
					AAC Asn					1755
					AGT Ser 470					1803
- •					CAT His					1851
					AGA Arg					1899
_					CAA Gln					1947
					TTC Phe					1995
_					CAA Gln 550					2043
					GTT Val					2091
					AGA Arg					2139
	_				AAG Lyb					2187

										GTC Val 615					CAG Gln 620	22	235
										ATT						22	283
										ACC Thr						23	331
										GAT Asp						23	379
										ATT					CCA Pro	24	127
										ATT 11e 695						24	175
										AAA Lys					λGλ λrg	25	23
										TTC Phe					CAA Gln	25	71
										GAT Asp				_		26	19
										CCG Pro						26	67
										ATG Met 775						27	15
										TCA Ser						27	63
ATA Ile	AGA Arg	CAG Gln	AAC Asn 800	AAT ABD	CAG Gln	CAT His	TTC Phe	AAT Asn 805	GAC Asp	ATA Ile	AAC ABn	CAG Gln	ATG Met 810	CTA Leu	G1 y	28	11
										CAA Gln						28	59
										AAC						29	07
AGC Ser 845	AAT Aen	AGT Ser	AAT Asn	AAC Asn	AGA Arg 850	GTT Val	GAT Asp	AAT Asn	CCA Pro	GAA Glu 855	GAT Asp	ACA Thr	GTG Val	GAT Asp	TCT Ser 860	29	55

			ATT													3003
Ser	Val	Asp	Ile	Thr 865	Ala	Phe	Tyr	Asp	Pro 870	Arg	Met	Ser	Ser	875	Ser	
				005					870					6/3		
															TAC	3051
Lys	Phe	Anb	Trp	Glu	Val	Ser	Lys		Hi B	Vel	yab	Pro		Ala	Tyr	
			880					885					890			
TCG	GTT	AAC	GTT	GCT	AGT	GAA	λλC	CGT	GTA	CTG	GAC	GAC	TTT	AAG	AAA	3099
Ser	Val		Val	Ala	Ser	Glu		λrg	Val	Leu	Asp		Phe	Lys	Lys	
		895					900					905				
GCA	TTT	CGC	GAA	λλG	AGA	λλλ	TAAC	TAC	TT J	ATTT1	CATI	C TO	CGAC	CAGA		3150
	Phe		Glu			Lys										
	910					915										
TTG	TAC	AT :	TTAC	TTTC	T G7	CCTC	TGAT	TC	ATA	TOT	ACAZ	TATA	ATT (	GAC	ATTTA	3210
																10
TAGI	ATAC	CAA 2	ATATA	CACC	A TO	CAATO	TATA	CA1	CCA	TATC	ACTI	CTCC	TA J	AAGA	CATCCC	3270
<b>ጉ</b> ጥጉባ	"TAAT	'AG '	TACAC	CCAT	T 1		2723	CAT	CATT	244	CTTC	ACTT	. J. C	*	AGCTT	3330
			nond	CON		DUUU	min	· CA	on I	, ALT	0.10	.no i	inc (	-m.	JAGC11	3330
ATT?	KTTA'	CC (	TTGC	TTTF	G A	TTTT	CCAA	GTC	CTAK	TTT	GTTI	TTTC	TA I	ACGC1	TTGCAA	3390
cerc	יש ייי	ירא ו		·	· ~ ~ ~	~~~~	ACCE	Cha	~~~	~ 7 3	acc.	TOTO	~~~ a	Per 8 er e	стстс	2450
	.n.c.i	CA /	10011	CIIC		JUCA	MUCA	GAI	CIIC	AAU.	ACC.	11010	.61 1	11411	CICIC	3450
AATG	CTGI	TC (	CACI	TTCA	T C	TCGI	CTGG	GAA	AAGI	CACC	GGTA	AGGG	CG			3500

### (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 915 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Het Leu Gln Phe Asn Thr Glu Asn Asp Thr Val Ala Pro Val Phe Pro 1 5 10 15

Het Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val Pro Leu Val Gln
20 25 30

Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu Pro Ile Val Phe 35 40 45

Leu Lys Gly Phe Glu Thr Asn Gly Leu Ser Glu Ile Ala Pro Ser Ile 50 55

Leu Arg Gly Ser Leu Ile Val Arg Val Leu Lys Pro Asn Lys Leu Lys 65 70 75 80

Ser Ile Ser Ile Thr Phe Lys Gly Ile Ser Arg Thr Glu Trp Pro Glu

Gly Ile Pro Pro Lys Arg Glu Glu Phe Ser Asp Val Glu Thr Val Val 100 105

Asn His Thr Trp Pro Phe Tyr Gln Ala Asp Asp Gly Met Asn Ser Phe 115 120 125

Thr Leu Glu His His Ser Ser Asn Asn Ser Ser Asn Arg Pro Ser Met 130 135 140

Ser 145		Glu	Авр	Tyr	Leu 150	Leu	Glu	Lye	Ser	Gly 155		Ser	Val	Tyr	11e 160
Fro	Pro	Thr	λla	Glu 165	Pro	Pro	Lys	Asp	Aen 170		Asn	Leu	Ser	Leu 175	
Ala	Tyr	Glu	<b>A</b> rg 180	λsn	Ser	Leu	Ser	Ser 185		Aen	Leu	Ser	Asn 190	Lys	Pro
Val	Ser	Ser 195	Asp	Val	Ser	His	<b>Asp</b> 200	двр	Ser	Lys	Leu	Leu 205		Ile	Gln
Lys	Thr 210	Pro	Leu	Pro	Ser	Ser 215	Ser	λrg	Àгд	Gly	Ser 220		Pro	Ala	Asn
Phe 225	His	Gly	yeu	Ser	Leu 230	Ser	Pro	His	Thr	Phe 235	Ile	Ser	yab	Leu	Phe 240
Thr	Lys	Thr		Ser .245	Asn	Ser	Gly	Ala	Thr 250	Pro	Ser	Pro	Glu	Gln 255	Glu
Asp	Asn	Tyr	Leu 260	Thr	Pro	Ser	Lys	Авр 265	Ser	Lys	Glu	Val	Phe 270	Ile	Phe
Àгд	Pro	Gly 275	Хвр	Tyr	lle	Tyr	Thr 280	Phe	Glu	Gln	Pro	11e 285	Ser	Gln	Ser
Tyr	Pro 290	Glu	Ser	Ile	Lys	Ala 295	Asn	Phe	Gly	Ser	Val 300	Glu	Tyr	Lys	Leu
Ser 305	Ile	Хвр	Ile	Glu	Arg 310	Phe	Gly	Ala	Phe	Lys 315	Ser	Thr	Ile	His	Thr 320
Gln	Leu	Pro	Ile	Lys 325	Val	Val	Arg	Leu	Pro 330	Ser	Авр	Gly	Ser	Val 335	Glu
Glu	Thr	Glu	340	lle	Ala	lle	Ser	Lys 345	Asp	Trp	Lys	Авр	Leu 350	Leu	His
Tyr	Asp	Val 355	Val	Ile	Phe	Ser	Lys 360	Glu	lle	Val	Leu	365	Ala	Phe	Leu
Pro	11e 370	Авр	Phe	His	Phe	375	Pro	Leu	Asp	Lys	Val 380	Thr	Leu	His	λrg
11e 385	λrg	Ile	Tyr	Leu	Thr 390	G] u	Ser	Het	Glu	Tyr 395	Thr	Сув	Asn	Ser	Asn 400
Gly	Asn	His	Glu	Lys 405	Ala	λrg	Arg	Leu	Glu 410	Pro	Thr	Lys	Lys	Phe 415	Leu
Leu	Ala	Glu	His 420	Asn	Gly	Pro	Lys	Leu 425	Pro	His	Ile	Pro	Ala 430	Gly	Ser
Asn	Pro	Leu 435	Lys	λla	Lys	Asn	Arg 440	Gly	Asn	Ile	Leu	Leu 445	увЬ	Glu	Lye
Ser	Gly 450	двр	Leu	Val	Aen	Lys 455	Авр	Phe	Gln	Phe	Glu 460	Val	Phe	Val	Pro
Ser 465	Lys	Phe	Thr	Asn	Ser 470	lle	Arg	Leu	His	Pro 475	Asp	Thr	λen	Tyr	<b>Asp</b> 480
Lys	Ile	Lys	Ala	His 485	His	Trp	Ile	Lys	11e 490	Су <b>в</b>	Leu	Àrg	Leu	Ser 495	Lys

Lys	Tyr	Gly	Авр 500	Asn	Arg	Lys	His	Ph <b>e</b> 505	Glu	lle	Ser	Ile	<b>Asp</b> 510	Ser	Pro
Ile	His	Ile 515	Leu	Asn	Gln	Leu	Сув 520		His	Ala	Asn	Thr 525	Leu	Leu	Pro
Ser	Tyr 530	Glu	Ser	His	Phe	Gln 535	Tyr	Сув	qaA	Glu	Авр 540	Gly	Asn	Phe	Ala
Pro 545	Ala	Ala	двр	Gln	Gln 550	λsn	Tyr	λla	Ser	His 555	His	Авр	Ser	λøn	11e 560
Phe	Phe	Pro	Lys	Glu 565	Val	Leu	Ser	Ser	Pro 570	Val	Leu	Ser	Pro	<b>Asn</b> 575	Val
Gln	Lys	Het	Asn 580	Ile	Arg	Ile	Pro	Ser 585	Asp	Leu	Pro	Val	Val 590	λrg	ABn
Arg	Ala	Glu 595	Ser	Val	Lys	Lys	5er 600	Lys	Ser	Авр	λsn	Thr 605	Ser	Lys	Lys
λæn	<b>хвр</b> 610	Gln	5er	Ser	<b>As</b> n	Val 615	Phe	Ala	Ser	Lys	Gln 620	Leu	Val	Ala	Asn
11e 625	Tyr	Lys	Pro	Nen	Gln 630	Ile	Pro	Arg	Glu	Leu 635	Thr	Ser	Pro	Gln	Ala 640
Leu	Pro	Leu	Ser	Pro 645	Il•	Thr	Ser	Pro	11e 650	Leu	yeu	Tyr	Gln	Pro 655	Leu
Ser	Asn	Ser	Pro 660	Pro	Pro	Asp	Phe	Asp 665	Phe	Asp	Leu	Ala	Lys 670	Arg	Gly
Ala	Ala	Asp 675	Ser	His	Ala	11e	Pro 680	Val	qeK	Pro	Pro	Ser 685	Tyr	Phe	Авр
Val	Leu 690	Lys	Ala	Двр	Gly	11e 695	Glu	Leu	Pro	Tyr	Tyr 700	Хвр	Thr	Ser	5er
Ser 705	Lys	Ile	Pro	Glu	Leu 710	Lys	Leu	Asn	Lys	Ser 715	λrg	Glu	Thr	Leu	Ala 720
Ser	Ile	Glu	Glu	Asp 725	Ser	Phe	уви	Gly	Trp 730	Ser	Gln	Ile	Двр	735	Leu
Ser	Asp	Glu	Авр 740	Asp	Asn	Asp	Gly	745	lle	Ala	Ser	Gly	Phe 750	λen	Phe
Lys	Leu	Ser 755	Thr	Ser	Ala	Pro	Ser 760	Glu	λen	Val	Asn	Ser 765	His	Thr	Pro
Ile	Leu 770	Gln	Ser	Leu	λen	Met 775	Ser	Leu	λвр	Gly	Arg 780	Lys	Lys	λsn	λrg
785	Ser	Leu	Hi∎	Ala	Thr 790	Ser	Val	Leu	Pro	Ser 795	Thr	Ile	Arg	Gln	neA 008
Asn	Gln	His	Phe	Asn 805	Авр	Ile	Asn	Gln	Het 810	Leu	Gly	Ser	Ser	Asp 815	Glu
Asp	Ala	Phe	Pro 820	Lys	Ser	Gln	Ser	Leu 825	naA	Phe	Asn	Lys	Lys 830	Leu	Pro
11	Leu	Lys 835	11e	Asn	Asp	Asn	Val 840	Ile	Gln	Ser	λεπ	Ser 845	Asn	Ser	Asn

Asn Arg Val Asp Asn Pro Glu Asp Thr Val Asp Ser Ser Val Asp Ile 850 855 860

Thr Ala Phe Tyr Asp Pro Arg Met Ser Ser Asp Ser Lys Phe Asp Trp 865 870 875 880

Glu Val Ser Lys Asn His Val Asp Pro Ala Ala Tyr Ser Val Asn Val 885 890 895

Ala Ser Glu Asn Arg Val Leu Asp Asp Phe Lys Lys Ala Phe Arg Glu 900 905 910

Lys Arg Lys 915

- (2) INFORMATION FOR SEQ ID NO:6:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2745 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: cDNA to mRNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (C) INDIVIDUAL ISOLATE: coding sequence of CNI
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATGCTCCAAT TCAATACAGA AAATGATACT GTAGCTCCAG TGTTTCCCAT GGAGCAAGAT 60 ATARATECAS CACCEGATEC CETCCCACE ETECAGACAA CAACACTACA AGECTTEGTA 120 ANGETTGEEG ANCECATAGT GTTTTTANAN GGATTTGANA CTANEGGAET GTCTGANATA 180 GCCCCCAGTA TCTTACGAGG ATCTCTTATC GTCAGGGTGT TGAAACCGAA TAAATTAAAA 240 AGTATATCGA TAACCTTCAA AGGAATATCC AGAACAGAGT GGCCGGAAGG TATACCACCG 300 ANGAGAGAAG AATTTTCAGA TGTTGAAACT GTTGTCAATC ACACATGGCC ATTTTATCAG 360 GCGGATGACG GCATGAATTC TTTCACCTTA GAACATCACA GCTCAAATAA TTCGTCCAAT 420 CGCCCATCTA TGAGCGATGA AGATTATCTA CTTGAAAAAA GCGGTGCTTC AGTATATATC 480 CCACCAACCG CTGAACCCCC TAAAGATAAT AGCAATCTAA GTCTGGATGC CTATGAGCGC 540 AACTCATTGT CATCCGATAA TTTGAGTAAC AAGCCAGTAT CAAGTGATGT TTCCCATGAC 600 GACAGTAAAC TGTTGGCTAT TCAAAAGACA CCATTACCAT CATCTAGTCG AAGAGGATCG 660 GTACCGGCAA ATTTTCACGG TAACTCTTTG TCACCTCATA CCTTCATATC TGATTTGTTC 720 ACANANACAT TCAGTANTAG TGGCGCTACT CCNAGTCCTG AGCNAGAGGA TAACTATCTT 780 ACACCATCCA AAGATTCTAA AGAAGTTTTT ATTTTTCGAC CGGGCGATTA TATTTACACT 840 TTTGAACAGC CAATATCGCA ATCTTATCCA GAAAGTATAA AAGCCAATTT TGGTTCCGTG 900 GAGTATAAAC TGTCAATAGA CATAGAGAGG TTTGGCGCAT TCAAATCAAC TATACATACT 960

CAATTA	CCCA	TCAAAGTCGT	AAGGCTTCCT	TCTGATGGAT	CCGTAGAAGA	GACTGAAGCT	1020
ATTGCA	TTTA	CCAAGGACTG	GAAAGATCTT	CTICATTATG	ACGTGGTAAT	TTTCTCGAAA	1080
GAGATO	GTTT	TGAATGCATT	TTTACCCATC	GATTTCCATT	TCGCTCCTCT	AGATAAAGTT	1140
ACTCTG	CATC	GTATTAGAAT	TTATCTAACA	GAGT <b>CTATGG</b>	AATACACTTG	TAATAGTAAT	1200
GGAAAT	CACG	AG <b>AA</b> GGCTCG	TAGATTAGAG	ССААСТАААА	AGTTTCTGTT	GGCTGAACAT	1260
AACGGT	CCTA	AACTGCCTCA	TATACCAGCT	GGTTCGAATC	CTTTGAAGGC	TAAAAATAGA	1320
GGGAAC	ATCC	TCTTGGATGA	AAAATCCGGC	GATCTAGTTA	ACAAAGATTT	TCAGTTCGAG	1380
GTGTTT	GTCC	CAAGCAAGTT	TACAAACAGT	ATACGGTTAC	ACCCTGATAC	AAATTATGAT	1440
AAAATC	AAAG	CCCACCATTG	GATAAAAATT	TGCCTTCGTC	TTTCCAAGAA	GTACGGGGAC	1500
AATAGA	AAAC	ATTTCGAAAT	AAGTATTGAT	TCTCCAATCC	ATATTTTAAA	TCAACTATGC	1560
TCACAC	GCGA	ATACTTTGCT	ACCGAGCTAC	GAGAGTCATT	TCCAGTATTG	TGATGAAGAT	1620
GGTAAT	TTCG	CACCAGCAGC	AGATCAACAA	AATTACGCAA	GTCATCATGA	TTCCAATATT	1680
TTCTTC	CCAA	AAGAAGTTCT	TTCGTCTCCC	GTTCTTTCAC	CTAACGTGCA	GAAGATGAAC	1740
ATTAGA	ATAC	CGTCTGATCT	TCCAGTAGTG	CGTAATAGAG	CTGAAAGCGT	AAAGAAAAGC	1800
AAGTCA	GATA	ATACCTCCAA	GAAGAATGAT	CAAAGTAGCA	ATGTCTTCGC	ATCCAAACAG	1860
CTGGTC	GCAA	ACATTTATAA	GCCCAATCAG	ATTCCAAGAG	AATTAACTTC	TCCTCAGGCG	1920
TTACCA	TAT	CGCCCATCAC	CTCACCAATT	CTCAATTACC	AACCATTATC	AAACTCCCCG	1980
CCTCCA	GATT	TTGATTTTGA	TCTAGCTAAG	CGCGGCGCAG	CCGATTCTCA	TGCTATTCCT	2040
GTGGAT	ССТС	CATCATATTT	TGATGTATTA	AAGGCCGATG	GGATTGAATT	GCCATACTAC	2100
GATACA	AGTT	CATCTAAAAT	TCCTGAACTA	AAACTAAACA	AATCTAGAGA	GACATTGGCC	2160
AGCATT	GAGG	AGGACTCATT	CAATGGTTGG	TCTCAAATTG	ATGACTTATC	CGACGAAGAT	2220
GACAAT	GATG	GCGATATAGC	ATCTGGTTTC	AACTTCAAGC	TGTCAACCAG	TGCTCCGAGT	2280
GAGAAC	GTTA	ATTCACACAC	TCCTATTTTG	CAGTCTTTAA	ACATGAGTCT	TGATGGGAGA	2340
XXXXXX	AATC	GTGCCAGTCT	<b>ACACGCAACA</b>	TCAGTGTTAC	CTAGTACAAT	AAGACAGAAC	2400
AATCAG	CATT	TCAATGACAT	AAACCAGATG	CTAGGCAGTA	GTGACGAAGA	TGCCTTTCCC	2460
AAAAGC	CAAT	CATTAAATTT	CAATAAGAAA	CTACCAATAC	TTAAAATTAA	TGATAACGTC	2520
ATACAA:	CAA	ACAGCAATAG	TAATAACAGA	GTTGATAATC	CAGAAGATAC	AGTGGATTCT	2580
TCAGTC	GATA	TTACAGCATT	TTATGATCCA	AGAATGTCAT	CAGATTCCAA	ATTTGATTGG	2640
GAGGTA	AGCA	AGAACCATGT	TGACCCAGCA	GCCTACTCGG	TTAACGTTGC	TAGTGAAAAC	2700
CGTGTA	CTGG	ACGACTTTAA	GAAAGCATTT	CGCGAAAAGA	GAAAA		2745

# (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: CNI-PRC-A
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

### CCAAAAAAG AGATCTCGGA TCAAAGTAGC

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- (2) INFORMATION FOR SEQ ID NO:8:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 30 base pairs(B) TYPE: nucleic acid

    - (C) STRANDEDNESS: mingle
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (C) INDIVIDUAL ISOLATE: CNI-PCR-B
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

## GGGTTTTTCA GTGTCGACGA TTCATAGATC

- (2) INFORMATION FOR SEQ ID NO:9:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1964 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNA1 coding sequence
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 286..1944
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
- GTTTGTTGCA TTTTGATATT CATCTATATC TATTTCAAAA TTTTTCATGT CATCGCCTCT

TGA	AACA	TGA	TTTA	TCCA	L LY	CTGA	AAAA	G AA	CGTA	CTAC	TGG	GAAA	CAA	AAGG	GAAAA	120
TGT	<b>KATA</b>	ATC	CTTT	ኢእፕር	ר דד	TTGA	ATCA	A GA	GGCA	TATT	TAT	<b>KAAA</b>	GAA	CGA	GCAAAC	180
CCT	<b>KATT</b>	TAT	TTGC	TTTA	TT A	AAGG	TATT	A TI	сааа	<b>KAA</b> D.	AGI	TITI	TTA	GATI	CTTTTT	240
TTT	TTGA	CGT	ATTA	GCTC	AG C	TGCC	ATAA	A AC	ACTO	TCAA	CG <b>C</b>		TG T et S			294
GAC Asp	TTG Leu 5	Asn	TCT Ser	TCA Ser	CGC	ATC Ile 10	Lys	ATC	ATT	AAA Lye	Pro 15	yeu	GAC Asp	TCT Ser	TAC	342
ATA Ile 20	Lys	GTT Val	GAC Asp	CGG	AAA Lys 25	Lys	GAT Asp	TTA	ACA Thr	AAA Lys 30	Tyr	GAA Glu	TTA	GAA Glu	λΑC λen 35	390
GGT	AAA Lys	GTA Val	ATT	TCT Ser 40	ACT	AAG Lys	GAC Asp	CGA Arg	TCC Ser 45	TAC Tyr	GCT Ala	TCT	GTA Val	Pro 50	Ala	438
ATA Ile	ACA Thr	GGA Gly	AAG Lys 55	ATA Ile	CCA Pro	AGT Ser	GAT Asp	GAG Glu 60	GAA Glu	GTA Val	TTC Phe	GAC Asp	TCC Ser 65	AAG Lys	ACG Thr	486
Gly	Leu	Pro 70	Asn	His	Ser	Phe	TTA Leu 75	Arg	Glu	His	Phe	Phe 80	His	Glu	Gly	534
Arg	Leu 85	Ser	Lys	Glu	Gln	Ala 90	ATA	Lys	Ile	Leu	<b>Хал</b> 95	Met	Ser	Thr	Val	582
100	Leu	Ser	Lys	Glu	Pro 105	Asn	CTA Leu	Leu	Lys	Leu 110	Lys	Ala	Pro	Ile	Thr 115	630
ATA Ile	TGT Cys	GGT	GAT Asp	ATT Ile 120	CAC His	GGG Gly	CAG Gln	TAT Tyr	TAT Tyr 125	GAT Asp	TTA Leu	TTG Leu	AAA Lys	CTG Leu 130	TTT Phe	678
GAA Glu	GTT Val	ej gec	GGT Gly 135	ABP	CCC Pro	GCC Ala	GAA Glu	ATC Ile 140	GAC ABP	TAT	TTA Leu	TTC Phe	TTG Leu 145	GGG Gly	GAT Asp	726
TAT Tyr	GTT Val	GAT Asp 150	усу Усу	CJA CCL	) Ala	TTC Phe	TCT Ser 155	TTT Phe	GAG Glu	TGT Cys	CTG Leu	ATT Ile 160	TAT Tyr	TTG Leu	TAC Tyr	774
TCC Ser	TTG Leu 165	AAG Lys	TTG Leu	AAT ABN	AAT Aen	TTA Leu 170	GGG	AGA Arg	TTT Phe	TGG Trp	ATG Met 175	CTA Leu	AGA Arg	GGT Gly	AAC Asn	822
CAT His 180	GAG Glu	TGT Cys	AAG Lys	CAC Him	TTG Leu 185	ACC Thr	TCT Ser	TAT Tyr	TTT Phe	ACT Thr 190	TTT Phe	AAG Lys	AAT Aen	GAG Glu	ATG Met 195	870
TTG Leu	CAC His	AAA Lys	TAC Tyr	GAT Asp 200	ATG Met	GAA Glu	GTT Val	TAC Tyr	GAT Asp 205	GCT Ala	TGC Cys	TGC Cy•	AGA Arg	TCA Ser 210	TTC Phe	918
AAT Asn	GTC Val	TTA Leu	CCA Pro 215	TT <b>λ</b> Leu	GCA Ala	GCT Ala	TTA Leu	ATG Met 220	AAC Aen	GGA Gly	CA <b>A</b> Gln	TAT Tyr	TTT Phe 225	TGT Cys	GTG Val	966

		_	Ile					Lys					Val		AAA Lys	1014
		Arg					Pro					Met			CTA Leu	1062
CTA Leu 260	Trp	GCC Ala	GAT	CCT Pro	GTC Val 265	Glu	AAT Asn	TAT	GAT Asp	GAT Asp 270	Ala	AGA Arg	GAT	Gly	AGC Ser 275	1110
									GTA Val 285						Gly	1158
									Ser					Lys	GCA Ala	1206
			Leu						CAC						GCG Gly	1254
									ACA Thr							1302
	Met								GAC Amp		Tyr					1350
									ATG Met 365							1398
									GAT Asp							1446
									GTT Val							1494
ATA Ile	TTA Leu 405	AAC	ATA Ile	TGT Cys	AGT Ser	GAG Glu 410	CAG Gln	GAA Glu	CTT Leu	GAC Asp	CCA Pro 415	GAA Glu	TCG Ser	GAA Glu	CCC Pro	1542
									AGA Arg							1590
									TCA Ser 445							1638
									AAG Ly≊							1686
						Val			GAA Glu							1734
									TTA Leu							1782

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_	 		 			CTA Leu					1830
			 			TTA Leu 525					187 <b>8</b>
	 	 	 			GAA Glu					1926
	 AAG Lys 550	 	 TGAT	[ጸጹል]	rc <b>t</b> :	CAT	rtta:	T			1964

### (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 553 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) HOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Het Ser Lys Asp Leu Asn Ser Ser Arg Ile Lys Ile Ile Lys Pro Asn 1 5 10 15

Asp Ser Tyr Ile Lys Val Asp Arg Lys Lys Asp Leu Thr Lys Tyr Glu 20 25 30

Leu Glu Asn Gly Lys Val Ile  $S \in r$  Thr Lys Asp Arg Ser Tyr Ala Ser 35 40

Val Pro Ala Ile Thr Gly Lys Ile Pro Ser Asp Glu Glu Val Phe Asp 50 55

Ser Lys Thr Gly Leu Pro Asn His Ser Phe Leu Arg Glu His Phe Phe 65 70 75 80

His Glu Gly Arg Leu Ser Lys Glu Gln Ala Ile Lys Ile Leu Asn Het 85 90 95

Ser Thr Val Ala Leu Ser Lys Glu Pro Asn Leu Leu Lys Leu Lys Ala 100 105 110

Pro Ile Thr Ile Cys Gly Asp Ile His Gly Gln Tyr Tyr Asp Leu Leu 115 120 125

Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Glu Ile Asp Tyr Leu Phe 130 140

Leu Gly Asp Tyr Val Asp Arg Gly Ala Phe Ser Phe Glu Cys Leu Ile 145 150 155 160

Tyr Leu Tyr Ser Leu Lys Leu Asn Asn Leu Gly Arg Phe Trp Met Leu 165 170 175

Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Ph Thr Phe Lys 180 185 190

Asn Glu Het Leu His Lys Tyr Asp Het Glu Val Tyr Asp Ala Cys Cys 195 200

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Arg	Ser 210		Asn	Val	Leu	Pro 215		Ala	Ala	Leu	Met 220		Gly	Gln	Туз
Phe 225	Сув	Val	His	Gly	Gly 230		Ser	Pro	Glu	Leu 235	Lys	Ser	Val	Glu	Asp 240
Val	λen	Lys	Ile	Авп 245	Arg	Phe	λrg	Glu	11e 250		Ser	λrg	Gly	Leu 25£	Het
Сув	Хвр	Leu	Leu 260		Ala	увр	Pro	Val 265	Glu	Asn	Tyr	увр	Asp 270	Ala	λrç
λвр	Gly	Ser 275		Phe	увр	Gln	Ser 280	Glu	λвр	Glu	Phe	Val 285	Pro	Asn	Sez
Leu	Arg 290	Gly	Сув	Ser	Phe	Ala 295	Phe	Thr	Phe	Lys	Ala 300	Ser	Сув	Lys	Phe
Leu 305	Lys	Ala	<b>As</b> n	Gly	Leu 310	Leu	Ser	Ile	Ile	Arg 315	Ala	His	Glu	Ala	Glr. 320
Авр	λla	Gly	Tyr	Arg 325	Met	Tyr	Lys	Asn	<b>Ав</b> п 330	Lys	Val	Thr	Gly	Phe 335	Pro
Ser	Leu	lle	Thr 340	Met	Phe	Ser	Ala	Pro 345	Asn	Tyr	Leu	Авр	Thr 350	Tyr	His
Asn	Lys	Ala 355	Ala	Val	Leu	Lys	Tyr 360	Glu	Glu	naA	Val	Het 365	λen	Ile	λrg
Gln	Phe 370	Hie	Met	Ser	Pro	His 375	Pro	Tyr	Trp	Leu	Pro 380	увь	Phe	Met	Asp
Val 385	Phe	Thr	Trp	Ser	Leu 390	Pro	Phe	Val	Gly	Glu 395	Lys	Val	Thr	Ser	Met 400
Leu	Val	Ser	Il•	Leu 405	Asn	Ile	Сув	Ser	Glu 410	Gln	Glu	Leu	ysb	Pro 415	Glu
Ser	Glu	Pro	Lys 420	Ala	Ala	Glu	Glu	Thr 425	Val	Lys	Ala	λrg	Ala 430	λen	λla
Thr	Lys	Glu 435	Thr	Gly	Thr	Pro	Ser 440	Авр	Glu	Lys	Ala	Ser 445	Ser	Ala	Ile
Leu	Glu 450	Хэр	Clu	Thr	Arg	Arg 455	Lys	Ala	Leu	λrg	<b>Asn</b> 460	Lys	Ile	Leu	λla
11e 465	λla	Lys	Val	Ser	Arg 470	Met	Phe	Ser	Val	leu 475	λrg	Glu	Glu	Ser	Glu 480
Lys	Val	Glu	Tyr	Leu 485	Lys	Thr	Met	λsn	Ala 490	Gly	Val	Leu	Pro	Arg 495	Gly
Ala	Leu	Ala	<b>Arg</b> 500	Gly	Thr	Glu	Gly	<b>Leu</b> 505	λsn	Glu	Thr	Leu	Ser 510	Thr	Phe
Glu	Lys	Ala 515	'nrg	Lys	Glu	Авр	Leu 520	Ile	nak	Glu	Lys	Leu 525	Pro	Pro	Ser
Leu	Ser 530	Glu	Val	Glu	Gln	Glu 535	Lys	Ile	Lys	Tyr	Tyr 540	Glu	Lys	Ile	Leu
Lye 545	Gly	Ala	Glu	Lys	Lys 550	Pro	Gln	Leu							

(2) INFORMATION P	FOR SEQ	ID	NO:11:
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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2353 base pairs
    (B) TYPE: nucleic acid
    (C) STRANDEDNESS: double

  - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNA2 coding sequence
- (ix) FEATURE:

  - (A) NAME/KEY: CDS (B) LOCATION: 262..2073
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

ATAGTCTATA ATACGTTTGA TACAGCTAGA TATCGCTAGC GCCAACATTG TCCCCCTCTC	60
TTGATCAATG CTTTTTTCG GCCCGAGACA AATGAGAAAA TGTCCTAAAA ATACCTTTCA	120
TCAAGACTCC TATTTTTCCT TAGAAAAAAC ATATATCCAA CTGGAACAGT ATTAAGCCAA	180
TTGCTACGAT ACAAACAAAA GGAGATATTC CTTCCCTCCC ATAGAGTCAC ACAGGAGCCA	240
GTACTTCTTC TTGAACCCGC A ATG TCT TCA GAC GCT ATA AGA AAT ACT GAG Het Ser Ser Asp Ala Ile Arg Asn Thr Glu 1 5 10	291
CAG ATA AAC GCC GCT ATT AAA ATT ATA GAA AAC AAA ACA GAG CGT CCG Gln Ile Asn Ala Ala Ile Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro 15 20 25	339
CAA TCG TCC ACA ACC CCT ATA GAT TCG AAG GCT AGT ACA GTT GCT GCT Gln Ser Ser Thr Thr Pro Ile Abp Ser Lyb Ala Ser Thr Val Ala Ala 30 35 40	387
GCT AAT TCC ACG GCC ACA GAA ACT TCC AGA GAC CTT ACA CAA TAT ACC Ala Asn Ser Thr Ala Thr Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr 45 50 55	435
CTA GAT GAC GGA AGA GTC GTA TCG ACA AAC CGC AGA ATA ATG AAT AAA Leu Asp Asp Gly Arg Val Val Ser Thr Asn Arg Arg Ile Met Asn Lys 60 65 70	483
GTG CCC GCC ATC ACG TCA CAT GTT CCT ACA GAT GAA GAG CTG TTC CAG Val Pro Ala Ile Thr Ser His Val Pro Thr Asp Glu Glu Leu Phe Gln 75 80 85 90	531
CCC AAT GGG ATA CCT CGT CAC GAA TTC CTA AGA GAT CAT TTC AAG CGC Pro Asn Gly Ile Pro Arg His Glu Phe Leu Arg Asp His Phe Lys Arg 95 100 105	579
GAG GGC AAA TTG TCG GCT GCG CAG GCG GCC AGG ATC GTT ACA CTT GCA Glu Gly Lys Leu Ser Ala Ala Gln Ala Ala Arg Ile Val Thr Leu Ala 110 115 120	627
ACG GAA CTC TTC AGC AAA GAA CCC AAC CTT ATA TCT GTT CCC GCC CCA Thr Glu Leu Phe Ser Lys Glu Pro Asn Leu Ile Ser Val Pro Ala Pro 125 130 135	675

		Val					Hie					Авр			AAG Lys	7 <b>2</b> 3
	Phe					Asp					Ser				TTG Leu 170	773
					Arg					Phe					TAT	819
				Lys					Asp					Leu	AGG Arg	867
								Thr							AAT Asn	915
															GAA Glu	963
														TAT Tyr	CTT Leu 250	1011
														GAC Asp 265	ATT Ile	1059
														ATG Het		1307
														TTG	GAT Asp	1155
														GTT Val	CCT Pro	1203
														AAC Asn		1251
		Gly	Сув	Ser	Tyr	Ala	Phe	Thr		λrg	Ala	λla	Сув	CAT His 345		1299
														GCT Ala		1347
GAC Aop	GCT Ala	GGT Gly 365	TAT Tyr	AGA Arg	ATG Het	TAC Tyr	AAA Lys 370	AAT Abn	ACC Thr	AAG Lys	ACT Thr	TTG Leu 375	GGC Gly	TTT Phe	CCC Pro	1395
														TAC Tyr		1443
									Asn					ATC 11e		1491

								TAT Tyr								1535
								GTT Val 435								1587
								ACT Thr								1635
								GGT Gly								1683
_								CAA Gln								1731
								GAA Glu								1779
								GTT Val 515								1827
								TTT Phe								1875
								AAT Asn								1923
								λGA λrg								1971
								CTG Leu								2019
								CAT Him 595								2067
AGC Ser		TAGA	(GAA)	GC T	CCTA	TTTC	OK 33	TGT)	CATA	CTT	CAAT	AAG	TAAG	TAAG	TT	2123
GCAT	TAAT	TA 1	CTAT	TTAG	A AG	CTAG	ATGC	TCC	TCAA	ATG	CACA	GAAT	CA 1	ATAG	CGTTT	2183
TATI	AGGT	CT G	TTCI	TTAI	T TT	AGTI	TIGI	TGA	TCTC	TAT	<b>GYY</b> C	GTAT	AT I	KTAT	TGCAA	2243
λλλΊ	AAAC	TT 1	TAAA	TATO	T AT	GGAT	CCTI	ACT	CAAT	TGT	ATAG	ACGT	TT I	TCAT	'AGGAG	2303
TGCA	LTAK	'AT G	GACA	CCAC	C TT	СТАА	TTGA	GCA	GAAG	CGG	TTCT	GAAT	TC			2353

# (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  (A) LENGTH: 604 amino acids
  (B) TYPE: amino acid
  (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Met Ser Ser Asp Ala Ile Arg Asn Thr Glu Gin Ile Asn Ala Ala Ile 1 10 15

Lys Ile Ile Glu Ash Lye Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro  $20 \hspace{1cm} 25 \hspace{1cm} 30$ 

Ile Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr 35 40 45

Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val 50 60

Val Ser Thr Asn Arg Arg Ile Met Asn Lys Val Pro Ala Ile Thr Ser 65 70 80

His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg 85 90 95

His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala 100 105 110

Als Gln Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys 115 120 125

Glu Pro Amn Lau Ile Ser Val Pro Ala Pro Ile Thr Val Cym Gly Amp 130 140

11e His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly 145 150 155 160

Amp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Amp Tyr Val Amp Arg 165 170 175

Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu 180 185 190

Asn Phe Asn Asp His Phe Trp Leu Leu Arg Gly Asn His Glu Cys Lys 195 200

His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu His Lys Tyr 210 215 220

Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro 225 230 235 240

Leu Ala Ala Leu Het Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile 245 250 255

Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe 260 265 270

Arg Glu Ile Pro Ser His Gly Leu Met Cys Asp Leu Leu Trp Ala Asp 275 280 285

Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu 290 295 300

Asp Il Val Asn Ser Lys Thr Met Val Pro His His Gly Lys Met Ala 305 310 315 320

Pro Ser Arg Asp Met Phe Val Pro Asn Ser Val Arg Gly Cys Ser Tyr 325 330 335

- Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu 340 345
- Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met 355 360 365
- Tyr Lys Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe 370 375 380
- Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu 385 390 395 400
- Lys Tyr Glu Asn Asn Val Met Asn Ile Arg Gln Phe Asn Met Thr Pro 405 415
- His Pro Tyr Trp Leu Pro Asp Phe Het Asp Val Phe Thr Trp Ser Leu 420 425 430
- Pro Phe Val Gly Glu Lys Val Thr Glu Met Leu Val Ala Ile Leu Asn 435 440 445
- Ile Cys Thr Glu Amp Glu Leu Glu Amn Amp Thr Pro Val Ile Glu Glu 450 455
- Leu Val Gly Thr Asp Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala 465 470 475 480
- Thr Prc Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu 485 490 495
- Asp Asp Glu His Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala Val 500 505 510
- Ala Lys Val Ser Arg Met Tyr Ser Val Leu Arg Glu Glu Thr Asn Lys 515 520 525
- Val Gln Phe Leu Lys Asp His Asn Ser Gly Val Leu Pro Arg Gly Ala 530 540
- Leu Ser Asn Gly Val Lys Gly Leu Asp Glu Ala Leu Ser Thr Phe Glu 545 550 560
- Arg Ala Arg Lys His Asp Leu Ile Asn Glu Lys Leu Pro Pro Ser Leu 565 570
- Asp Glu Leu Lys Asn Glu Asn Lys Lys Tyr Tyr Glu Lys Val Trp Gln 580 585
- Lys Val His Glu His Asp Ala Lys Asn Asp Ser Lys 595 600
- (2) INFORMATION FOR SEQ ID NO:13:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 812 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full

# CNB1 coding sequence

# (ix) FEATURE:

(A) NAME/KEY: CDS (B) LOCATION: 54..104

# (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 181..652

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

(XI) DEPOLACE PROCESTITION. DEG ID NO. 13.	
ACTTGGTANC TCANTGGTGA TCAGAATCCA TAGAAGCATT TTTATTTCTT AAA ATG Het 1	56
GGT GCT GCT CCT TCC AAA ATT GTG GAT GGT CTT TTA GAA GAT ACA AAT Gly Ala Ala Pro Ser Lya Ile Val Aap Gly Leu Leu Glu Aap Thr Aan 5 10 15	104
TGTATGTACA CTTCGGAGTG AGGAAAAGAA AGAAAGGGGA AATTAACCGA TTTTACTAAC	164
ACTGACACTT TGAACA GTT GAT AGA GAT GAA ATT GAA AGG TTA AGG AAG Val Amp Arg Amp Glu Ile Glu Arg Leu Arg Lym 1 5 10	213
AGA TTC ATG AAA TTA GAT AGA GAT AGC TCA GGG TCT ATT GAT AAA AAT Arg Phe Het Lys Leu Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn 15 20 25	261
GAA TTT ATG AGC ATT CCT GGC GTT TCG TCA AAC CCT CTT GCT GGA CGT Glu Phe Het Ser lie Pro Gly Val Ser Ser Amn Pro Leu Ala Gly Arg 30 35 40	309
ATA ATG GAG GTT TTC GAT GCT GAT AAT AGT GGG GAC GTG GAT TTT CAA  Ile Het Glu Val Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln 45 50 55	357
GAG TTC ATC ACA GGA TTA TCC ATT TTC AGT GGG CGT GGG TCC AAG GAC Glu Phe lle Thr Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp 60 65 70 75	405
GAA AAG TTA AGA TTC GCC TTC AAA ATC TAC GAC ATT GAC AAG GAC GGT Glu Lys Leu Arg Phe Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly 80 85	453
TTC ATA TCC AAT GGT GAG TTG TTC ATC GTG TTG AAG ATT ATG GTA GGT Phe Ile Ser Asn Gly Glu Leu Phe Ile Val Leu Lys Ile Het Val Gly 95 100 105	501
TCT AAT CTG GAC GAT GAA CAG CTG CAA CAG ATA GTA GAT AGG ACG ATA Ser Amn Leu Amp Amp Glu Gln Leu Gln Gln Ile Val Amp Arg Thr Ile 110 115 120	549
GTG GAA AAC GAT AGC GAC GGC GAC GGA CGT TTA AGT TTC GAG GAG TTT Val Glu Asn Asp Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe 125 130 135	597
AAG AAT GCT ATC GAA ACC ACA GAA GTG GCC AAG AGT CTG ACA TTG CAA Lys Asn Ala Ile Glu Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln 145 150 155	645
TAC GATGTGTAAG ACTAGGGGAC ACTTCATTCA TTTATGGTAT GCCAATATTT Tyr Asp	698

TTAAGAAAAG AAGAATAATA CGCGATATTG TTTTTTAAGG AAGGAACGCA CACTCGCCCA 758
GTTAGAGTGC TGATGATATA TACATATATA TATGTATATG TAACAAACAA 1AAG 812

- (2) INFORMATION FOR SEQ ID NO:14:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 17 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr 1 5 10 15

Asn

- (2) INFORMATION FOR SEQ ID NO:15:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 157 amino acids
    - (B) TYPE: amino acid
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
- Val Amp Arg Amp Glu Ile Glu Arg Leu Arg Lym Arg Phe Met Lym Leu
- Amp Arg Amp Ser Ser Gly Ser Ile Amp Lym Amn Glu Phe Het Ser Ile 20 25 30
- Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Met Glu Val Phe 35 40
- Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln Glu Phe Ile Thr Gly 50 60
- Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Amp Glu Lys Leu Arg Phe 65 70 80
- Als Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn Gly 85 90 95
- Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly Ser Asn Leu Asp Asp 100 105 110
- Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile Val Glu Asn Asp Ser 115 120 125
- Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe Lys Asn Ala Ile Glu 130 140
- Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp 145 155

(2) INFORMATION	FOR	SEQ	ID	NO:16:
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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 524 base pairs (B) TYPE: nucleic scid (C) STRANDEDNESS: double

  - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
   (C) INDIVIDUAL ISOLATE: coding sequence of CNB1
- (ix) FEATURE:

  - (A) NAME/KEY: CDS (B) LOCATION: 1..524

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

				GAT Asp 10				ACA Thr	48
				TTA Leu				AAA Lys	96
				GAT Asp					144
				GCT Ala					192
				GAT Asp					240
				TCC Ser 90					288
	-	 	 	 AAG Lys	_	 -			336
				ATG Het					384
				AGG Arg					432
				GAG Glu					480
				ACA Thr 170			GT		524

### (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 174 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Het Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr

Asn Phe Asp Arg Asp Glu Ile Glu Arg Leu Arg Lys Arg Phe Met Lys
20 25 30

Amp Arg Amp Ser Ser Gly Ser Ile Amp Lym Amn Glu Phe Met Ser

Ile Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Met Glu Val

Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln Glu Phe Ile Thr

Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp Glu Lys Leu Arg 85 90 95

Pho Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn 100 105 110

Gly Glu Leu Phe Ile Val Leu Lys Ile Het Val Gly Ser Aen Leu Asp

Asp Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile Val Glu Asn Asp

Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe Lys Asn Ala Ile

Glu Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp 165 170

### (2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1812 base pairs

  - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
  - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: DNA fragment containing CNAldeltaC coding sequence
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 286..1812

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTT	ጥር ጥጥ	CCB	***	· ጉ ሞ አ	TH (	· » ~~~		C Th	<b>ምም</b> ም ር		Trans	*TTCB	ምርጥ	C) T C	CCCCCC	
GII	1011	GCA	1111	GAIA	.11 (	AICI	WIWI	CIA	1111	ሌለሌክ	. 111	1107	101	CATC	GCCTCT	60
TGA	AACA	TGA	ATTT	TCCA	AT T	CTGA	AAAA	G AA	CGTA	CTAC	TGG	GAAA	CAA	AAGG	GAAAAA	120
TGT	AATA	ATC	CITI	AATG	TT T	TTGA	ATCA	A GA	GGCA	TTAT	TAT	ሊአልአ	GAA	CGAA	GCAAAG	180
CCI	TTAA	TAT	TTGC	TTTA	TT A	AAGG	TATT	A TT	CAAA	GAAA	AGT	TTTT	TTA	GATT	CITTII	240
TTT	TTGA	CGT	<b>AT</b> TA	GCTC	AG C	TGCC	ATAA	A AC	ACTC	TCAA	ccc			CG A er L		294
							Lys				CCT Pro 15	Asn			TAC Tyr	342
	Lys					Lys					TAC					390
											GCT Ala					438
											TTC Phe					486
_											TTC Phe					534
											AAT Asn 95					582
											Lys					630
											TTA Leu					678
											TTA Leu					726
											CTG Leu					774
											ATG Met 175					822
											TTT Phe					870
											TGC Cys					918

									naK	GGA						966
										GTA Val						1014
_										GGT Gly						1062
										CAT Asp 270						1 <b>110</b>
										CCT Pro						1158
										TGC Cys						1206
	Gly									GAA Glu						1254
										GCC						1302
										ACA Thr 350					_	1350
										AAC Aøn						1398
										TTT Phe		λвр				1446
										ACT Thr						1494
ATA Ile	TTA Leu 405	λ»n	ATA Ile	TGT Cys	AGT Ser	GAG Glu 410	CAG Gln	GAA Glu	CTT Leu	GAC Asp	CCA Pro 415	GAA Glu	TCG Ser	GAA Glu	CCC Pro	1542
AAA Lys 420	GCT Ala	GCG Ala	GAG Glu	GAG Glu	ACT Thr 425	GTA Val	AAG Lys	GCA Ala	AGA Arg	GCA Ala 430	AAC Asn	GCA Ala	ACT Thr	AAG Lyb	GAG Glu 435	1590
ACC Thr	GGC Gly	ACC Thr	CCA Pro	TCT Ser 440	GAT Asp	GAA Glu	AAG Lys	GCG Ala	TCA Ser 445	TCA Ser	GCG Ala	ATA Ile	TTA Leu	GAA Glu 450	GAT Asp	1638
GAA Glu	ACC Thr	CGA Arg	λGλ λrg 455	AAG Lys	GCT Ala	TTG Leu	AGA Arg	AAT ABN 460	AAG Lys	ATA Ile	TTA Lou	GCT Ala	ATT Ile 465	GCT Ala	AAA Lys	1686
GTT Val	TCA Ser	AGA Arg 470	ATG Met	TTT Phe	TCG Ser	GTG Val	CTA Leu 475	AGA Arg	GAA Glu	GAG Glu	AGC Ser	GAA Glu 480	AAA Lys	GTG Val	GAA Glu	1734

TAT TTG AAA ACT ATG AAT GCC GGT GTC TTA CCT CGT GGT GCT CTA GCT 1782 Tyr Leu Lys Thr Met Asn Ala Gly Val Leu Pro Arg Gly Ala Leu Ala CGT GGG ACT GAA GGT TTG AAT GAA ACG CTA 1812 Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu

### (2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 509 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19: Het Ser Lya Asp Leu Asn Ser Ser Arg Ile Lys Ile Ile Lys Pro Asn Asp Ser Tyr Ile Lys Val Asp Arg Lys Lys Asp Leu Thr Lys Tyr Glu 20 25 30 Leu Glu Asn Gly Lys Val Ile Ser Thr Lys Asp Arg Ser Tyr Ala Ser Val Pro Ala Ile Thr Gly Lys Ile Pro Ser Asp Glu Glu Val Phe Asp Ser Lys Thr Gly Leu Pro Asn His Ser Phe Leu Arg Glu His Phe Phe 65 70 75 80 His Glu Gly Arg Leu Ser Lys Glu Gln Ala Ile Lys Ile Leu Asn Het 85 90 95 Ser Thr Val Als Leu Ser Lys Glu Pro Asn Leu Leu Lys Leu Lys Ala 100 Pro Ile Thr Ile Cys Gly Asp Ile His Gly Gln Tyr Tyr Asp Leu Leu Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Glu Ile Asp Tyr Leu Phe 130 140 Leu Gly Amp Tyr Val Amp Arg Gly Ala Phe Ser Phe Glu Cym Leu Ile

Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu Hie Lye Tyr Asp Met Glu Val Tyr Asp Ala Cys Cys

Tyr Leu Tyr Ser Leu Lys Leu Asn Asn Leu Gly Arg Phe Trp Het Leu

Arg Ser Phe Amn Val Leu Pro Leu Ala Ala Leu Met Amn Gly Gln Tyr

Phe Cys Val His Gly Gly Ile Ser Pro Glu Leu Lys Ser Val Glu Asp

Val Asn Lys Ile Asn Arg Ph Arg Glu Ile Pro Ser Arg Gly Leu Met 245 250

- Cys Asp Leu Leu Trp Ala Asp Pro Val Glu Asn Tyr Asp Asp Ala Arg
- Asp Gly Ser Glu Phe Asp Gln Ser Glu Asp Glu Phe Val Fro Asn Ser
- Leu Arg Gly Cys Ser Phe Ala Phe Thr Phe Lys Ala Ser Cys Lys Phe
- Leu Lys Ala Asn Gly Leu Leu Ser lle Ile Arg Ala His Glu Ala Gln
- Asp Ala Gly Tyr Arg Met Tyr Lys Asn Asn Lys Val Thr Gly Phe Pro 325 330 335
- Ser Leu Ile Thr Het Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr His
- Asn Lys Ala Ala Val Leu Lys Tyr Glu Glu Asn Val Het Asn Ile Arg
- Gln Phe His Het Ser Pro His Pro Tyr Trp Leu Pro Asp Phe Het Asp
- Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Ser Het
- Leu Val Ser Ile Leu Asn Ile Cys Ser Glu Gln Glu Leu Asp Pro Glu
- Ser Glu Pro Lys Ala Ala Glu Glu Thr Val Lys Ala Arg Ala Asn Ala
- Thr Lys Glu Thr Gly Thr Pro Ser Asp Glu Lys Ala Ser Ser Ala Ile
- Leu Glu Asp Glu Thr Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala 455
- Ile Ala Lys Val Ser Arg Het Phe Ser Val Leu Arg Glu Glu Ser Glu
- Lys Val Glu Tyr Leu Lys Thr Het Asn Ala Gly Val Leu Pro Arg Gly 490
- Ala Leu Ala Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu 500
- (2) INFORMATION FOR SEQ ID NO: 20:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1767 base pairs
    - (B) TYPE: nucleic scid
    - (C) STRANDEDNESS: double (D) TOPOLOGY: unknown
  - (ii) MOLECULE TYPE: DNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: DNA fragment containing CNA2deltaC coding sequence
  - (ix) FEATURE:

- (A) NAME/KEY: CDS
  (B) LOCATION: 262..1767

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:20:	
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								_								
ATA	GTCT	ATA	ATAC	GTTT	GA T	ACAG	CTAG	A TA	TCGC	TAGC	GCC	AACA	TTG	TCCC	CCTCTC	60
TTG	ATCA	ATG	CTTT	TTTT	C <b>G</b> 6	CCCG	AGAC	A AA	TGAG	<b>A</b> AA <b>A</b>	TGT	CCTA	AAA	ATAC	CTTTCA	120
TCA	<b>A</b> GAC	TCC	TATT	TTTC	CT T	AGAA	aaaa	C AT	TATA	CCAA	CTG	GAAC	AGT	ATTA	AGCCAA	180
TTC	CTAC	GAT	ACAA	ACAA	AA G	GAGA	TATT	с ст	TCCC	тсс <b>с</b>	ATA	GAGT	CAC	ACAG	GAGCCA	240
GTA	CTTC	TTC	TTGA	ACCC	GC A					GCT Ala 5					_	291
										AAC						339
										GCT Ala				Ala		387
										GAC Asp						435
										CGC						483
										GAT Asp 85						531
										AGA						579
										AGG Arg						627
										ATA Ile						675
										TAC Tyr						723
						_	_			ACA Thr 165				-	_	771
										TTT Phe						819
										CAT His						867

			Glu					Thr					Phe		TAK .	915
		Leu										Lys			GAA Glu	963
TCG Ser 235	Phe	AAC ABn	AAC Asn	TTG Leu	CCC Pro 240	Leu	GCT Ala	GCG Ala	TTA Leu	ATG Met 245	ysu	GGA Gly	CAG Gln	TAT	CTT Leu 250	1011
					Ile					Asn					ATT	1059
AAC ABD	AAC Asn	CTA Leu	AAT ABD 270	AGA Arg	TTC Phe	AGG Arg	GAG Glu	ATT Ile 275	CCC Pro	TCT Ser	CAT His	GGC Gly	CTG Leu 280	ATG Met	TCT Cys	1107
						CC <b>G</b> Pro									GAT Asp	1155
						GAC Asp 305										1203
						CCT Pro										1251
GTA Val	AGG Arg	GGC	TGT Cys	TCA Ser 335	TAT Tyr	GCC Ala	TTC Phe	ACG Thr	TAT Tyr 340	CGT Arg	GCT Ala	GCG Ala	TGC Cys	CAT His 345	TTT Phe	1299
						TTG Leu										1347
						TAC Tyr										1395
						AGT Ser 385										1443
						AAA Lys										1491
						CAC His										1539
						CCA Pro										1587
						ATC Ile										1635
						TTA Leu 465										1683

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GCT GGT ANG TCG GAN GCN ACT CCN CAN CCN GCC ACT TCG GCG TCG CCT 1731 Ala Gly Lys Ser Glu Ala Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro 480 485

AAA CAT GCT TCC ATT TTA GAT GAC GAA CAT CGA AGG Lys His Ala Ser Ile Leu Asp Asp Glu His Arg Arg 495 500

1767

- (2) INFORMATION FOR SEQ ID NO:21:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 502 amino acids(B) TYPE: amino acid

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Met Ser Ser Asp Ala Ile Arg Asn Thr Glu Gln Ile Asn Ala Ala Ile

Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro

Ile Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr

Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val
50 60

Val Ser Thr Asn Arg Arg Ile Het Asn Lys Val Pro Ala Ile Thr Ser 65 70 75 80

His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg

His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala

Ala Gln Ala Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys

Glu Pro Asn Leu Ile Ser Val Pro Ala Pro Ile Thr Val Cys Gly Asp

Ile His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly

Asp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg

Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu 185

Asn Phe Asn Asp His Phe Trp Leu Leu Arg Gly Asn His Glu Cys Lys 200

His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu His Lys Tyr

Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro

Leu Ala Ala Leu Met Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile

72

Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe 265 Arg Glu Ile Pro Ser His Gly Leu Het Cys Asp Leu Leu Trp Ala Asp Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu Amp Ile Val Amn Ser Lym Thr Het Val Pro Him Him Gly Lym Het Ala Pro Ser Arg Asp Het Phe Val Pro Asn Ser Val Arg Gly Cys Ser Tyr 330 Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met 355 360 365 Tyr Lys Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu Lys Tyr Glu Asn Asn Val Het Asn Ile Arg Gln Phe Asn Het Thr Pro His Pro Tyr Trp Leu Pro Asp Phe Het Asp Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Glu Het Leu Val Ala Ile Leu Asn

Ile Cys Thr Glu Asp Glu Leu Glu Asn Asp Thr Pro Val Ile Glu Glu

Leu Val Gly Thr Asp Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala

Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu

Asp Asp Glu His Arg Arg 500

#### (2) INFORMATION FOR SEQ ID NO: 22:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid

  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (C) INDIVIDUAL ISOLATE: G4-PCR-A

WO 96/12806

## PCT/US95/13580

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

GCCCTATCGT GCACTCACCG ACGC

24

- (2) INFORMATION FOR SEQ ID NO:23:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 24 base pairs

    - (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (C) INDIVIDUAL ISOLATE: G4-PCR-B
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GTGAAGGCCC TACTGAGCCA GGAG

#### IT IS CLAIMED:

- A polypeptide composition comprising a polypeptide effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin.
  - 2. A composition of claim 1, wherein the polypeptide composition contains a calcineurin interacting (CNI) polypeptide.
- 3. A composition of claim 2, wherein the polypeptide composition contains a polypeptide having an amino acid sequence selected from the group consisting of sequences represented by SEQ ID NO:2 and SEQ ID NO:5.
- A composition of claim 2, wherein the polypeptide has an amino acid sequence of
   between 15 and 915 amino acids in length.
  - 5. A composition of claim 2, wherein the polypeptide composition contains a polypeptide having an amino acid sequence comprising the c-terminal 306 amino acids of a CNI protein.

20

- 6. An isolated nucleic acid having a sequence encoding a polypeptide of any of claims 1-5.
- A nucleic acid of claim 6, wherein the nucleic acid has a sequence selected from
   the group consisting of nucleic acid sequences represented by SEQ ID NO:3 and SEQ ID NO:6.
  - 8. A method of identifying a small molecule immunosuppressant compound, comprising
- constructing a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains an (A) subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI polypeptide,

contacting the cell with a small molecule, and

identifying the small molecule as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins.

9. A method of claim 8, wherein the cell is a yeast cell.

5

- 10. A method of claim 8, wherein one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain.
- 10 11. A method of claim 8, wherein the subunit of calcineurin is selected from the group consisting of yeast CNA1 and yeast CNA2.
  - 12. A method of claim 8, wherein the subunit of calcineurin is an "A" subunit of human calcineurin.

- 13. A method of claim 8, wherein the CNI polypeptide is yeast CNI polypeptide.
- 14. A method of claim 8, wherein the CNI polypeptide is yeast CNIc polypeptide.
- 20 15. A method of claim 8, wherein the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell.
- 16. A yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where said mutation prevents expression of a
   functional calcineurin-interacting polypeptide from said genomic copy.

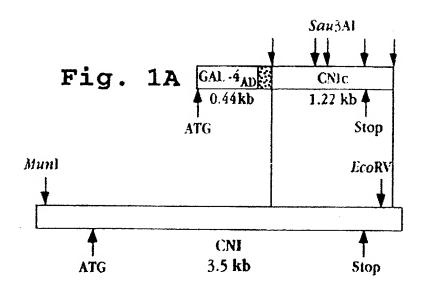
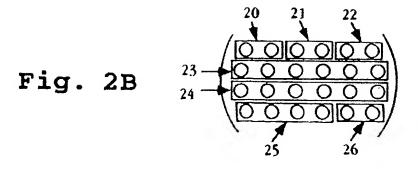


Fig. 1B





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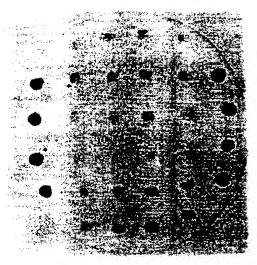


Fig. 3A

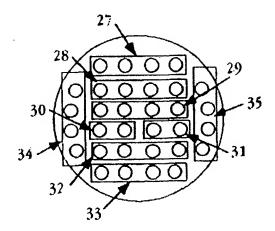
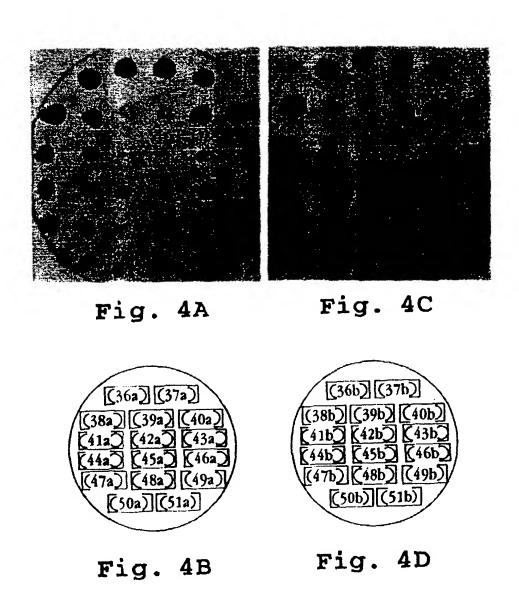


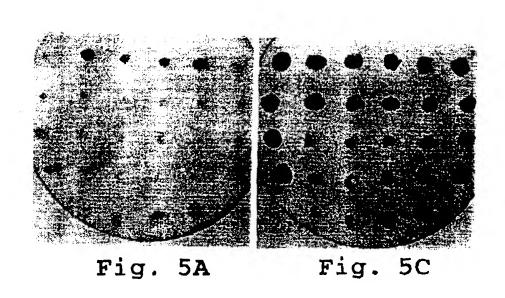
Fig. 3B

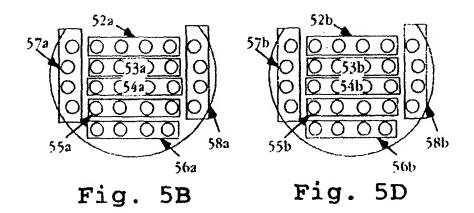
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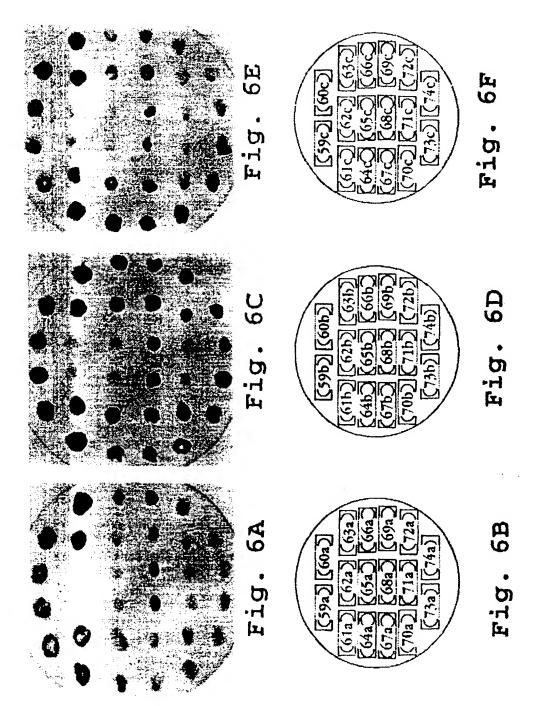
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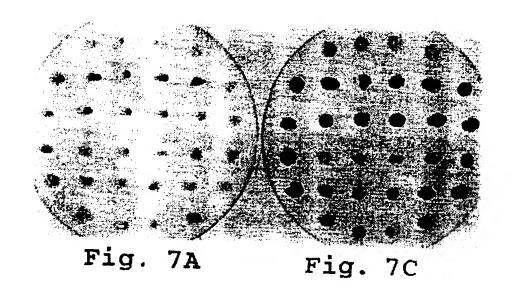


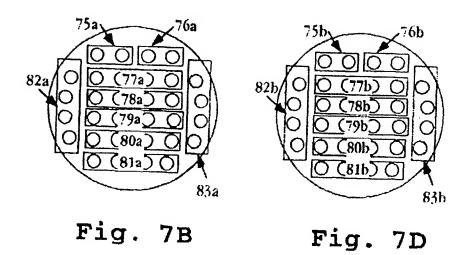
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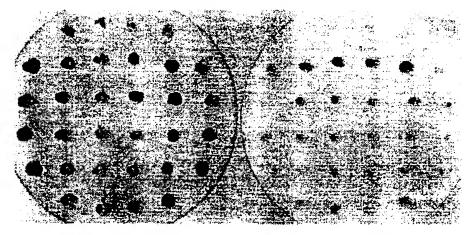


Fig. 7E

Fig. 7G

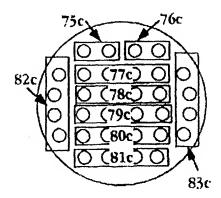


Fig. 7F

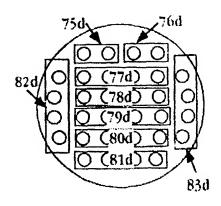


Fig. 7H

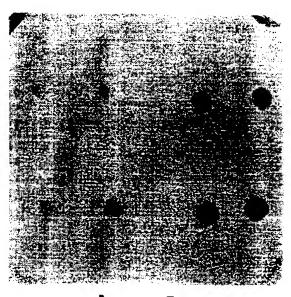


Fig. 8A

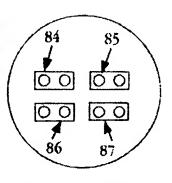


Fig. 8B



Fig. 9A



Fig. 9C

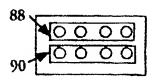


Fig. 9B

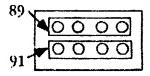
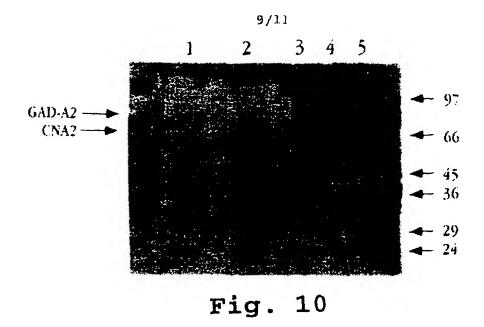
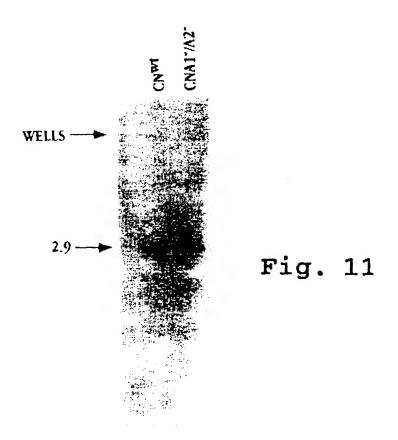


Fig. 9D

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Fig. 12

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Fig. 13

# INTERNATIONAL SEARCH REPORT Termahonal Application No.

1 PCT/US 95/13580

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Electronic d	ata base consulted during the international search (name of data b	ase and, where practical, search terms used	)		
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		<del></del>		
Category *	Citation of document, with indication, where appropriate, of the	relevant passages	Relevant to claim No.		
X	Swissprot Database entry YK01_YE Accession number P36117; June 01 Duesterhoeft A. et al.: 'Hypothet kD protein in YPT52-GCN3 interge region.'	,1994 ical 102.5	1-5		
X	Emfun Database entry Scykr021w Accession number Z28246; May 10, Duesterhoeft A. et al.:'S. cerev chromosome XI reading frame ORF	risiae	6,7		
P <b>,</b> X	MOLECULAR BIOLOGY OF THE CELL, 5 (SUPPL.). 1994. 141A., HUANG L ET AL 'A novel protein that interacts with calcineurin in vivo' see abstract 818		1,2,6, 8-15		
		-/			
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	DOON) DOCUMENTS CONSIDERED TO BE RELEVANT			
Category *	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.	
A	CURR BIOL, 2 (1). 1992. 18-20., CYERT M S 'IMMUNOSUPPRESSANTS HIT THE TARGET' see the whole document			